

Plants as source of new therapies for endometriosis: a review of preclinical and clinical studies

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BACKGROUND: Given the disadvantages and limitations of current endometriosis therapy, there is a progressive increase in studies focusing on plant-derived agents as a natural treatment option with the intention of achieving high efficiency, avoiding adverse effects and preserving the chance for successful pregnancy. The heterogeneity of these studies in terms of evaluated agents, applied approaches and outcomes illustrates the need for an up-to-date summary and critical view on this rapidly growing field in endometriosis research.

OBJECTIVE AND RATIONALE: This review provides a comprehensive overview of plant-derived agents and natural treatment strategies that are under preclinical or clinical investigation and critically evaluates their potential for future endometriosis therapy.

SEARCH METHODS: An English language PubMed literature search was performed using variations of the terms 'endometriosis', 'natural therapy', 'herb/herbal', 'plant', 'flavonoid', 'polyphenol', 'phytochemical', 'bioactive', 'Kampo' and 'Chinese medicine'. It included both animal and human studies. Moreover, the Clinicaltrials.gov database was searched with the term 'endometriosis' for clinical trials on plant-derived agents. No restriction was set for the publication date.

OUTCOMES: Natural therapies can be assigned to three categories: (i) herbal extracts, (ii) specific plant-derived bioactive compounds and (iii) Chinese herbal medicine (CHM). Agents of the first category have been shown to exert anti-proliferative, anti-inflammatory, anti-angiogenic and anti-oxidant effects on endometrial cells and endometriotic lesions. However, the existing evidence supporting their use in endometriosis therapy is quite limited. The most studied specific plant-derived bioactive compounds are resveratrol, epigallocatechin-3-gallate, curcumin, puerarin, ginsenosides, xanthohumol, 4-hydroxybenzyl alcohol, quercetin, apigenin, carnosic acid, rosmarinic acid, wogonin, baicalein, parthenolide, andrographolide and cannabinoids, with solid evidence about their inhibitory activity in experimental endometriosis models. Their mechanisms of action include pleiotropic effects on known signalling effectors: oestrogen receptor- α , cyclooxygenase-2, interleukin-1 and -6, tumour necrosis factor- α , intercellular adhesion molecule-1, vascular endothelial growth factor, nuclear factor-kappa B, matrix metalloproteinases as well as reactive oxygen species (ROS) and apoptosis-related proteins. Numerous studies suggest that treatment with CHM is a good choice for endometriosis management. Even under clinical conditions, this approach has already been shown to decrease the size of endometriotic lesions, alleviate chronic pelvic pain and reduce postoperative recurrence rates.

WIDER IMPLICATIONS: The necessity to manage endometriosis as a chronic disease highlights the importance of identifying novel and affordable long-term safety therapeutics. For this purpose, natural plant-derived agents represent promising candidates. Many of these agents exhibit a pleiotropic action profile, which simultaneously inhibits fundamental processes in the pathogenesis of endometriosis, such as proliferation, inflammation, ROS formation and angiogenesis. Hence, their inclusion into multimodal treatment concepts may essentially contribute to increase the therapeutic efficiency and reduce the side effects of future endometriosis therapy.

Key words: endometriosis / natural therapy / herb / plant / flavonoid / polyphenol / phytochemical / bioactive / Chinese herbal medicine

Introduction

Endometriosis is a common oestrogen-dependent gynecological disease, which is characterised by the presence of endometrial tissue outside the uterine cavity. The condition affects 5–10% of women of reproductive age and is usually associated with inflammation, severe pelvic pain and infertility (Burney and Giudice, 2012; Zondervan *et al.*, 2018). Moreover, endometriosis is a leading cause of miscarriage (Lessey *et al.*, 2013; Santulli *et al.*, 2016) and implantation failure (Margalioth *et al.*, 2006; Tomasetti *et al.*, 2006).

The eutopic endometrium of endometriosis patients differs from that of disease-free women and exhibits intrinsic characteristics that stimulate cell survival outside the uterine cavity (Liu and Lang, 2011). This supports the 'retrograde menstruation theory' of Sampson, postulating that eutopic endometrium gives rise to ectopic endometriotic lesions (Sampson, 1927). Pathological alterations in the endometrium of endometriosis patients may further explain the increased infertility rates associated with the disease. In fact, besides poor oocyte quality,

women with endometriosis exhibit decreased uterine receptivity and abnormal expression of endometrial biomarkers (Kao *et al.*, 2003; Vassiliadis *et al.*, 2005; Liu and Lang, 2011).

The formation and survival of endometriotic lesions at ectopic sites is crucially dependent on fundamental biological processes, including proliferation, apoptosis, inflammation and angiogenesis (Nasu *et al.*, 2009; Hey-Cunningham *et al.*, 2013; Laganà *et al.*, 2019) (Fig. 1). Cell proliferation is increased in newly developing endometriotic lesions when compared to eutopic endometrium (Rosa-e-Silva *et al.*, 2010). In addition, endometrial tissue from women with endometriosis is more resistant to apoptosis (Meresman *et al.*, 2000; Dmowski *et al.*, 2001; Szymanowski, 2007; Reis *et al.*, 2013; Roshangar *et al.*, 2013; Aznaurova *et al.*, 2014). This imbalance between proliferation and cell death promotes uncontrolled tissue growth. Endometriosis is also associated with inflammation in both the eutopic endometrium and the peritoneal cavity, characterised by altered humoral immunological function and enhanced numbers of activated peritoneal immune cells (Lousse *et al.*, 2010, 2012). This not only contributes to the

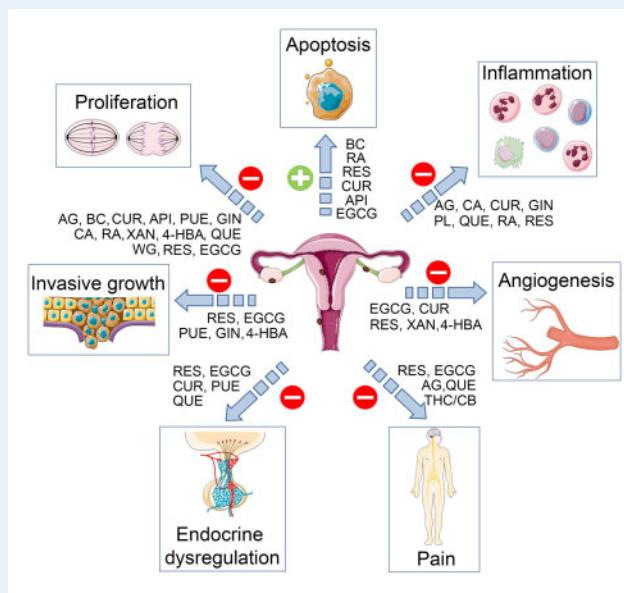


Figure 1. Targeting of dysregulated physiological processes in endometriosis by plant-derived bioactive compounds. Several physiological processes are dysregulated in endometriosis. Increased proliferation, invasion, inflammation and angiogenesis, resistance to apoptosis and dysregulation of steroid signalling contribute to lesion growth and hyperalgesia. As discussed in this review, several of these processes can be inhibited ('-' symbol) or promoted ('+' symbol; apoptosis) by plant-derived bioactive compounds. See text and Table II for details. AG, andrographolide; API, apigenin; BC, baicalein; CA, carnosic acid; CUR, curcumin; EGCG, epigallocatechin-3-gallate; GIN, ginsenosides; 4-HBA, 4-hydroxybenzyl alcohol; PL, parthenolide; PUE, puerarin; QUE, quercetin; RA, rosmarinic acid; RES, resveratrol; THC/CB, cannabinoids; WG, wogonin; XAN, xanthohumol.

persistence of ectopic endometriotic tissue against immunological defence mechanisms but also affects endometrial receptivity and pregnancy outcomes (Tomassetti *et al.*, 2006; Capobianco and Rovere-Querini, 2013). Another important hallmark in the pathogenesis of endometriosis is angiogenesis (Becker and D'Amato, 2007; Laschke and Menger, 2018). The ingrowth of new microvessels into ectopic endometrial tissue is a major prerequisite for its survival and growth (Groothuis *et al.*, 2005; Taylor *et al.*, 2009; Cho *et al.*, 2012). Accordingly, anti-angiogenic therapy has been shown to inhibit the development of experimental endometriosis (Laschke and Menger, 2007; Ricci *et al.*, 2011).

There is a close interaction of steroid hormones, the immune system and angiogenic pathways in the regulation of the aforementioned cellular processes in endometriosis (Gazvani and Templeton, 2002; Greene *et al.*, 2016). The peritoneal fluid from patients with endometriosis contains high levels of vascular endothelial growth factor (VEGF), which is released by both endometriotic lesions and peritoneal macrophages (Takehara *et al.*, 2004; Bourlev *et al.*, 2006). It has been further reported that VEGF induces cyclooxygenase-2 (COX-2) expression in endometrial and endometriotic cells (Attar and Bulun, 2006; Attar *et al.*, 2009; Bulun *et al.*, 2015). Oestrogen and COX-2, in

turn, enhance the expression of VEGF and matrix metalloproteinases (MMPs), which are also involved in angiogenesis. This vicious cycle finally maintains elevated concentrations of prostaglandin E₂ (PGE₂), a known mediator of active inflammation and pain in endometriotic tissue (Attar and Bulun, 2006) (Fig. 1).

The induction of hypoestrogenemia by combined oral contraceptives, progestins or gonadotropin releasing hormone agonists (GnRHs) is currently the most common pharmacological approach for the therapy of endometriosis. However, this type of treatment is unsatisfactory due to its side effects. Another option is the surgical removal of endometriotic lesions, which, however, is associated with high recurrence rates of up to 50% within 5 years of surgery (Guo, 2009; Guo and Martin, 2019).

Hence, it is of major importance to establish improved strategies for the management of endometriosis. To achieve this, an increasing number of studies focus on plant-derived agents as a natural treatment option with the intention of achieving high efficiency, avoiding adverse effects and preserving the chance for successful pregnancy. The present review provides a comprehensive overview of these agents and natural treatment strategies that are currently under preclinical or clinical investigation and critically evaluates their potential for future endometriosis therapy.

Methods

Articles were selected for relevance and quality from the English language PubMed literature database. The search was performed using variations of the terms 'endometriosis', 'natural therapy', 'herb/herbal', 'plant', 'flavonoid', 'polyphenol', 'phytochemical', 'bioactive', 'Kampo' and 'Chinese medicine'. It included both animal and human studies. Moreover, the Clinicaltrials.gov database was searched with the term 'endometriosis' for clinical trials on plant-derived agents. No restriction was set for the publication date.

Herbal Extracts

Phytomedicine has been practiced worldwide since ancient times to treat or alleviate human disease. Herbal extracts are highly complex multi-component mixtures. Hence, synergistic actions of their individual components may be highly beneficial (Zhou *et al.*, 2016). Accordingly, they have also been evaluated in different *in vitro* and *in vivo* models of endometriosis.

Pueraria flower extract

Pueraria lobata, also known as 'Kudzu', is a perennial leguminous plant consumed as a functional food in East Asia countries. Puerarin, one of the major isoflavones of the plant, is anti-oxidative and anti-inflammatory and inhibits aromatase activity in endometrial cells (Li *et al.*, 2008; Chen *et al.*, 2018a). Referring specifically to the herbal extract, the effects of pueraria flower extract (PFE) have been assessed on experimental endometriosis both *in vitro* and *in vivo*. It was found that PFE reduces MMP-2 and MMP-9 mRNA and protein levels in endometriotic cells and suppresses their adhesion and migration (Kim *et al.*, 2017). Moreover, mice, which were orally treated with PFE for

5 weeks beginning 1 week before the inoculation of endometrial fragments, exhibited a reduced number of newly developing endometriotic lesions when compared to vehicle-treated controls (Kim et al., 2017). These results suggest that PFE has potential as a therapy for preventing and treating endometriosis.

Hexane extract of aged black garlic

Garlic (*Allium sativum*) is a plant commonly used in many cultures of the world. It is one of the most widely researched medicinal plants that has been used as spice, food and medicine for over four thousand years. Extensive research has been carried out on the beneficial properties of garlic and its effects on various pathologies (Thomson and Ali, 2003). Moreover, different formulations of garlic have been developed. One of the most useful garlic preparations is aged black garlic. Several studies have shown that the extracts of aged black garlic exhibit a broad pharmacological effect spectrum, including pro-apoptotic, anti-oxidant and anti-cancer activity (Thomson and Ali, 2003; Park et al., 2014). Recently, Kim et al. (2013) suggested that hexane extract of aged black garlic (HEABG) to be effective in the prevention and treatment of endometriosis. They found that HEABG inhibits cell proliferation and cell cycle progression of tumour necrosis factor (TNF)- α -activated human endometriotic stromal cells isolated from patients with ovarian endometrioma through inhibition of the ERK and JNK signalling pathways (Kim et al., 2013). In addition, treatment of endometriotic stromal cells with HEABG potently suppressed TNF- α -induced intercellular adhesion molecule (ICAM)-1 and vascular adhesion molecule (VCAM)-1 expression by inhibiting the activation of nuclear factor-kappa B (NF- κ B) and nuclear activator protein-1 transcription factors (Kim et al., 2013). The latter mechanism may inhibit the recruitment of immune cells into endometriotic lesions and, thus, contribute to reduce disease-associated inflammation.

Uncaria tomentosa extract

Cat's claw is a specific plant from the Amazon rain forest. It is named after its leafy pattern that resembles the claws of a cat. There are several species of cat's claw, but the two most common varieties used are *Uncaria tomentosa* and *U. guianensis*. *U. tomentosa* is clearly the preferred variety because of its higher alkaloid content and the ease of standardisation (Erowele and Kalejaiye, 2009). Diseases which have been successfully treated with *U. tomentosa* include rheumatoid arthritis, prostatitis and cancer (Della Valle, 2017). The plant exerts anti-oxidant and anti-inflammatory effects mediated by the inhibition of PGE₂. In addition, it can alter cell cycle progression by inducing apoptosis (Neto et al., 2011). In rats with surgically induced endometriosis, *Uncaria tomentosa* extract (UTE) reduced the growth of ectopic lesions when compared to controls (Neto et al., 2011).

Açaí extract

Açaí, the fruit of an Amazonian plant, is a berry grown on the palm tree *Euterpe oleracea*. In the last years, research on the açaí fruit has focused on its anti-oxidant, anti-inflammatory and anti-tumoural properties (Ulbricht et al., 2012). Machado et al. (2016) evaluated the therapeutic potential of açaí extract (AE) on the growth of endometriotic lesions in rats. They found that AE significantly induces the regression and decreases the size of endometriotic lesions. Moreover, the treated

lesions exhibited reduced levels of VEGF, MMP-9, COX-2, PGE₂ and nitric oxide when compared to controls (Machado et al., 2016).

Viburnum opulus extract

Viburnum opulus, also called European cranberry bush or guelder rose, is widespread in Europe, North and Central Asia as well as North Africa. The *V. opulus* fruits are used as food plant, herbal teas and medicinal plant to treat a wide range of illnesses (Polka et al., 2019). In animal studies, *Viburnum opulus* extract (VOE) protected the male reproductive system from damage caused by taxanes (Sanözkan et al., 2017). In addition, it was found that daily oral treatment with VOE significantly reduces the volume of endometriotic lesions in rats (Saltan et al., 2016). Moreover, VOE-treated animals exhibited significantly lower peritoneal levels of TNF- α , VEGF and interleukin (IL)-6. Of note, these effects may be partially attributed to the chlorogenic acid compound of VOE (Saltan et al., 2016).

Silymarin extract

Milk thistle (*Silybum marianum*) is a medicinal plant from the Asteraceae family native in Southern Europe, Australia, North and South America, Northern Africa and some parts of Asia. Milk thistle has been used since ancient times for the treatment of liver diseases and for increasing milk production in lactating mothers (Soleimani et al., 2019). Silymarin extract (SE), the major constituent of milk thistle, is a mixture of flavonolignan and flavonoid polyphenolic compounds isolated from *S. marianum*, widely known for its potent anti-oxidant, anti-inflammatory, anti-proliferative and pro-apoptotic effects (Di Costanzo and Angelico, 2019). Accordingly, SE has been proposed as an anti-cancer and anti-metastatic natural compound (Hosseiniabadi et al., 2019). In rats with surgically induced endometriosis, the total serum anti-oxidant activity was significantly higher in SE-treated animals when compared to controls (Jouhari et al., 2018). Two recent studies further demonstrated that SE induces the regression of endometriotic lesions in a rat model (Jouhari et al., 2018; Nahari and Razi, 2018). Additional molecular and histopathological analyses revealed an up-regulation of ERK1/2 expression and inhibition of angiogenesis, resulting in increased apoptosis and fibrosis within SE-treated endometriotic lesions (Nahari and Razi, 2018). Nevertheless, more studies are necessary to clarify whether SE is also appropriate for the treatment of human endometriosis. Clinical trials indicate that SE is safe in humans at therapeutic doses and well tolerated even at a high dose (>1500 mg/day) (Soleimani et al., 2019). However, the compound exhibits low solubility in water, low bioavailability and poor intestinal absorption. Therefore to improve its bioavailability at the site of absorption, the use of nanotechnology appears to be a promising method, allowing a sustained release and increased the therapeutic action of the active herbal extract (Di Costanzo and Angelico, 2019).

Calligonum comosum extract

Calligonum comosum is a leafless perennial shrub widespread in sand dunes. It grows naturally in the North African deserts, the desert sands of the Middle East, Pakistan and both Central and Eastern Arabia (Soliman et al., 2018). *Calligonum comosum* is useful in traditional folkloric medicine for the treatment of abnormally heavy or prolonged menstruation (Tahmasebi et al., 2018) and exhibits anti-

Table I Medicinal plants and herbal extracts evaluated in experimental models of endometriosis.

| Herbal medicine | Model | Action/mechanism | Reference |
|-------------------------------------|--|---|---|
| Pueraria flower extract | Human endometriotic cell lines (11Z and 12Z) Balb/c mice | ↓ Cell migration and adhesion ↓ MMP-2 and MMP-9 levels ↓ Endometriotic lesion size ↓ Cell proliferation ↓ Cell cycle progression ↓ ERK and JNK pathways ↓ TNF- α -induced ICAM-1 and VCAM-1 expression by inhibiting the activation of NF- κ B and AP-1 | Kim et al. (2017) Kim et al. (2017) Kim et al. (2013) |
| Hexane extract of aged black garlic | Primary human endometriotic stromal cells isolated from patients with ovarian endometrioma | | |
| Uncaria tomentosa extract | Wistar rats | ↓ Endometriotic lesion size | Neto et al. (2011) |
| Açaí extract | Sprague-Dawley rats | ↓ Endometriotic lesion size ↓ VEGF, MMP-9 and COX-2 ↓ PGE ₂ , VEGF and NO levels | Machado et al. (2016) |
| Viburnum opulus extract | Sprague-Dawley rats | ↓ Endometriotic lesion size ↓ TNF- α , VEGF and IL-6 levels | Saltan et al. (2016) |
| Silymarin extract | Wistar rats Wistar rats | ↓ Endometriotic lesion size ↓ Endometriotic lesion size ↑ ERK1/2 expression ↓ Angiogenesis ↑ Apoptosis ↑ Fibrosis | Jouhari et al. (2018) Nahari and Razi (2018) |
| Calligonum comosum extract | Balb/c mice | ↓ Endometriotic lesion size ↓ Vascularisation ↓ Cell proliferation ↓ Immune cell infiltration | Kiani et al. (2019) |

AP-1, nuclear activator protein-1; COX-2, cyclooxygenase-2; ICAM-1, intercellular adhesion molecule-1; IL-6, interleukin-6; MMP, matrix metalloproteinase; NF- κ B, nuclear factor-kappa B; NO, nitric oxide; PGE₂, prostaglandin E₂; PGF₂- α , prostaglandin F₂- α ; TNF- α , tumour necrosis factor- α ; VCAM-1, vascular cell adhesion molecule-1; VEGF, vascular endothelial growth factor.

inflammatory, anti-oxidant and anti-cancer activity (Badria et al., 2007; Abdel-Sattar et al., 2014; Abdo et al., 2015). Recent findings indicate that *Calligonum comosum* extract (CCE) may also be beneficial for the treatment of endometriosis. After administration of CCE to mice with surgically induced endometriosis, an inhibition of growth and cyst formation of developing endometriotic lesions was observed. This was associated with reduced vascularisation, cell proliferation and immune cell infiltration within the lesions (Kiani et al., 2019).

Summary

As summarised in Table I, the findings of the aforementioned basic studies indicate that several herbal extracts show potential for future use in endometriosis therapy. This is most probably due to their broad action profile, which includes anti-proliferative, anti-oxidant, anti-inflammatory, anti-angiogenic and pro-apoptotic activity. Future studies have to clarify now whether these beneficial effects observed under experimental conditions are also reproducible in endometriosis patients. For this purpose, it will be necessary to establish standardised protocols for the good manufacturing practice (GMP)-compliant generation of well-defined plant extracts. This is a major prerequisite for the systematic evaluation of their efficiency and safety in human endometriosis therapy.

Specific Plant-derived Bioactive Compounds

Specific plant-derived bioactive compounds with evidence about their inhibitory activity on endometriosis include resveratrol, epigallocatechin-3-gallate (EGCG), curcumin, puerarin, ginsenosides, xanthohumol, 4-hydroxybenzyl alcohol (4-HBA), quercetin, apigenin, carnosic acid, rosmarinic acid, wogonin, baicalein, parthenolide, andrographolide and cannabinoids (Fig. 1).

Resveratrol

Resveratrol is a natural phytoalexin polyphenol synthesised by plants for the protection against fungal infections. High resveratrol levels are detected in grapes, wine, berries and nuts (Rauf et al., 2018; Dull et al., 2019). Besides its cardioprotective effects, resveratrol exhibits anti-cancer properties, as indicated by its ability to suppress the proliferation of a wide variety of tumour cells (Kundu and Surh, 2008). In addition, resveratrol exhibits an anti-oxidant, anti-inflammatory and anti-angiogenic activity (Kundu and Surh, 2008; Dull et al., 2019).

Several *in vitro* studies have analysed the effect of resveratrol on endometrial and endometriotic cells (Fig. 2). Bruner-Tran et al. (2011) demonstrated that resveratrol dose-dependently suppresses the invasiveness of human endometrial stromal cells into Matrigel®

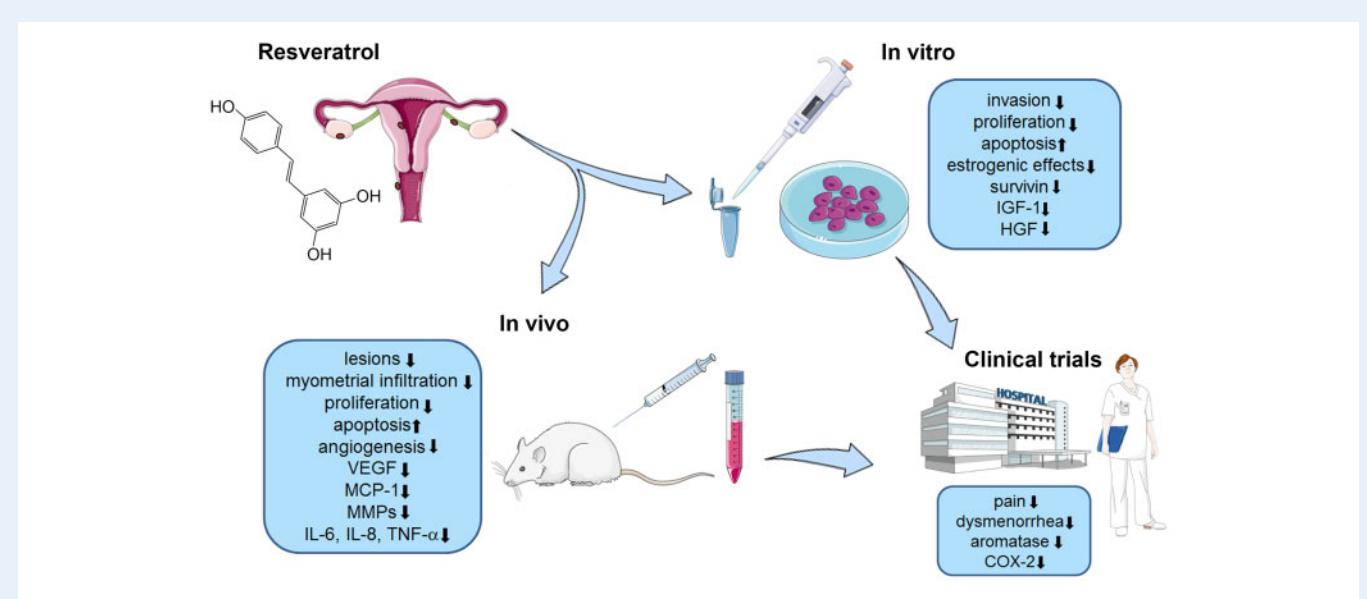


Figure 2. Resveratrol as a potential therapeutic in endometriosis. Preclinical *in vitro* studies on the polyphenol resveratrol have demonstrated an inhibitory effect on invasive growth and cell proliferation as well as a pro-apoptotic function. These effects can be partially attributed to an inhibitory effect on the expression of survivin, IGF-1 and HGF, while its effects on oestrogen are more complex. *In vivo* models of endometriosis confirm the anti-invasive and pro-apoptotic role as well as an inhibitory impact on angiogenesis, which is linked to a reduction in the expression of pro-inflammatory, pro-angiogenic and pro-invasive factors. A limited number of small clinical trials on the combination of oral contraceptives with resveratrol have revealed possible positive effects with respect to pain relief and decreased dysmenorrhea, however, one clinical trial failed to demonstrate positive effects. See text for details.

(Bruner-Tran et al., 2011). Moreover, it inhibits cell proliferation and induces apoptosis in primary epithelial endometrial cell cultures from endometriosis and control women (Ricci et al., 2013). Taguchi et al. (2016) found that resveratrol suppresses survivin expression and enhances tumour necrosis factor-related apoptosis-inducing ligand-induced apoptosis in endometriotic stromal cells from ovarian endometriosis (Taguchi et al., 2016). *In vitro* effects of resveratrol have also been studied by Amaya et al. (2014) using the Ishikawa endometrial cell line. They reported that resveratrol alone has a weak oestrogen activity while in the presence of the steroid hormone it can act as either an oestrogen agonist or antagonist at low and high concentrations, respectively. Moreover, resveratrol may reduce endometrial cell proliferation through ER- α (Amaya et al., 2014). Another study recently showed that resveratrol treatment of eutopic and ectopic endometrial stromal cells from endometriosis patients reduces their expression of insulin-like growth factor-1 (IGF-1) and hepatocyte growth factor (HGF), both important factors involved in the growth, proliferation and angiogenesis of endometriotic lesions (Arablou et al., 2019).

In line with these promising *in vitro* results, several *in vivo* studies reported beneficial effects of resveratrol on endometriotic lesions in mice (Fig. 2). In a first report, human endometrial tissue was injected into the peritoneal cavity of oestrogen-stimulated nude mice, which either received resveratrol or vehicle by daily gavage (Bruner-Tran et al., 2011). Resveratrol treatment reduced the fraction of animals developing endometriosis and the number of lesions as well as their total volume per mouse. In addition, resveratrol-treated lesions exhibited an increased apoptotic activity. Of note, these effects were observed at rather high resveratrol doses of 6 mg/mouse, which are not

reasonable for human administration (Bruner-Tran et al., 2011). Even higher doses of 60 mg/mouse were used in another immunocompromised murine model of endometriosis (Amaya et al., 2014), reporting an anti-oestrogenic and anti-proliferative effect of the compound on human endometrial xenografts. However, beneficial effects on experimental endometriosis have also been induced in mice by applying markedly lower resveratrol doses ranging between 10 and 40 mg/kg per day (Ricci et al., 2013; Rudzitis-Auth et al., 2013). These effects included the suppression of cell proliferation and angiogenesis as well as the stimulation of apoptosis, resulting in a reduced engraftment and growth of endometriotic lesions (Ricci et al., 2013; Rudzitis-Auth et al., 2013).

In a rat model of endometriosis, Ergenoglu et al. (2013) reported a significant reduction in implant size and considerable histological changes in endometriotic foci following a 14-day treatment with 10 mg/kg i.m. resveratrol. This was associated with decreased levels of VEGF and monocyte chemoattractant protein (MCP)-1 in the peritoneal fluid of the animals (Ergenoglu et al., 2013). In a similar model, Cenksoy et al. (2015) observed a regression of endometriotic lesions and lower VEGF and MCP-1 levels after a 21-day oral administration of resveratrol in a dose of 60 mg/kg. Of interest, they further compared the therapeutic efficiency of resveratrol with conventional GnRH α treatment and found that resveratrol was as effective as leuprolide acetate (Cenksoy et al., 2015). Yavuz et al. (2014) treated rats with intraperitoneal doses of 1 or 10 mg/kg/day resveratrol, resulting in reduced cell proliferation rates and endometriotic implant volumes. Moreover, the activities of superoxide dismutase and glutathione peroxidase as well as the levels of malonyl dialdehyde and catalase increased in the serum

and tissue of the animals when compared to controls. Based on these data, the authors suggested that resveratrol suppresses the development of surgically induced rat endometriotic implants probably due to its anti-oxidative properties (Yavuz *et al.*, 2014). In addition, Bayoglu Tekin *et al.* (2015) showed that resveratrol is a potent inhibitor of endometriosis-associated inflammation and angiogenesis. They detected reduced levels of the pro-inflammatory cytokines IL-6, IL-8 and TNF- α in the peritoneal fluid of resveratrol-treated rats and a lower expression of MMPs and VEGF in endometriotic implants when compared to controls (Bayoglu Tekin *et al.*, 2015).

Although resveratrol is the most extensively studied stilbene and various preclinical studies indicate its effectiveness in the treatment of endometriosis, clinical endometriosis trials with resveratrol are limited. Maia *et al.* (2012) conducted a clinical study including 42 women with endometriosis. Of these, 16 patients received oral contraceptives alone, whereas 26 patients were treated with a combination of oral contraceptives and resveratrol. It was found that the expression of aromatase and COX-2 was reduced in the eutopic endometrium of patients treated with combined therapy when compared to the control group (Maia *et al.*, 2012). An additional study of the same authors conducted in 12 patients with endometriosis, who had failed to obtain pain relief during the use of combined oral contraceptives, further reported a clinical benefit of resveratrol. The addition of 30 mg resveratrol to their daily oral contraceptive regimen caused a significant reduction in pain scores, with 82% of patients reporting complete resolution of dysmenorrhea and pelvic pain after 2 months of therapy (Maia *et al.*, 2012). Taken together, these results indicate that resveratrol potentiates the effects of oral contraceptives in the management of endometriosis-associated dysmenorrhea, by further reducing aromatase and COX-2 expression in the endometrium, although this should be confirmed in larger trials (Fig. 2). More recently, Mendes da Silva *et al.* (2017) conducted a randomised clinical trial involving 44 women with laparoscopic diagnosis of endometriosis to evaluate the effectiveness of resveratrol in the management of endometriosis associated with monophasic contraceptive pills. However, they found that the dose of 40 mg/day resveratrol did not improve endometriosis-related symptoms when compared to placebo-treated controls (Mendes da Silva *et al.*, 2017).

Epigallocatechin-3-gallate

EGCG is a flavonoid found both in black, but especially in green tea. In recent years, the compound has been considered for the treatment of different types of cancer based on its anti-oxidant, anti-angiogenic and anti-proliferative effects (Ju *et al.*, 2007; Shanmugam *et al.*, 2011; Singh *et al.*, 2015). The first study evaluating the effect of EGCG in the treatment of experimental endometriosis was performed in Syrian golden hamsters (Laschke *et al.*, 2008). EGCG suppressed the proliferation and VEGF expression of isolated hamster endometrial stromal and glandular cells. *In vivo*, EGCG inhibited angiogenesis, blood perfusion and growth of endometriotic lesions within dorsal skinfold chambers (Laschke *et al.*, 2008). Xu *et al.* (2009) subcutaneously transplanted eutopic endometrium from patients with endometriosis into immunodeficient mice, which were intraperitoneally treated with EGCG for 2 weeks. This treatment significantly suppressed the development of endometriotic lesions by inhibition of angiogenesis, as indicated by a decreased microvessel density and reduced mRNA levels of VEGF

within the lesions. In addition, the lesions contained higher numbers of apoptotic cells when compared to controls (Xu *et al.*, 2009). In a follow-up study, Xu *et al.* (2011) further reported that EGCG selectively suppresses VEGFC/VEGF receptor-2 (VEGFR-2) expression and signalling in experimental endometriosis *in vivo* and in endothelial cells *in vitro* (Xu *et al.*, 2011). In addition, they evaluated a prodrug of EGCG (pro-EGCG, EGCG octaacetate) with improved stability and bioavailability (Wang *et al.*, 2013). This prodrug exhibited a stronger anti-oxidant activity than EGCG and, thus, represents a promising compound for endometriosis therapy (Wang *et al.*, 2013). Ricci *et al.* (2013) were the first demonstrating the effectiveness of oral EGCG administration in a murine model of endometriosis. They showed that oral gavage of EGCG for 4 weeks inhibits the development of endometriotic lesions by diminishing cell proliferation and increasing apoptosis. Moreover, EGCG treatment reduced the number of established lesions per mouse (Ricci *et al.*, 2013). Using epithelial endometrial cells from human biopsies, it was further shown that EGCG also suppresses cell proliferation and induces cell death in cells of human origin (Ricci *et al.*, 2013). A more recent study revealed that EGCG inhibits cell proliferation, migration, invasion and collagen gel contraction of endometrial and endometriotic stromal cells *in vitro* (Matsuzaki and Darcha, 2014). This was associated with a significantly decreased expression of genes known to be involved in fibrogenesis. Interestingly, the anti-proliferative effect of EGCG was significantly more pronounced in endometriotic stromal cells than in endometrial stromal cells from patients with endometriosis. These findings are in accordance with other studies observing that EGCG more strongly suppresses cell proliferation of diseased cells than of their normal cell counterparts (Chen *et al.*, 2011a). Moreover, it was shown that EGCG prevents the progression of fibrosis within endometriotic lesions in a xenograft model (Matsuzaki and Darcha, 2014).

Of interest, the Chinese University of Hong Kong is currently recruiting for a randomised interventional clinical trial on 185 prospective endometrioma patients, who will be given high-purity EGCG (SUNIPHENON EGCG[®]; 400 mg, twice per day) or a placebo for 3 months prior to their planned surgery. Change in endometriotic lesion size (by MRI) is listed as a primary outcome, whereas pain scores, quality of life, change in lesion growth (by biopsy), change in neovascularisation and monitoring of adverse effects are listed as secondary outcomes (Wang, 2020a).

Curcumin

Curcumin is a polyphenolic compound found in turmeric, which is derived from the rhizomes of the plant *Curcuma longa* Linn (Bisht *et al.*, 2010). The anti-inflammatory and cytoprotective properties of curcumin have been widely reported (Bisht *et al.*, 2010; Kim *et al.*, 2012). The anti-inflammatory effect of curcumin on endometriosis was investigated by Kim *et al.* (2012) using primary endometriotic stromal cells obtained from ovarian endometrioma. They observed that this compound suppresses ICAM-1 and VCAM-1 gene and protein expression as well as the secretion of IL-6, IL-8 and MCP-1 by inhibiting the TNF- α -induced activation of NF- κ B. However, it had no significant effect on the viability of human endometriotic stromal cells (Kim *et al.*, 2012). In another *in vitro* study, Zhang *et al.* (2013a) found that curcumin is able to inhibit the proliferation of endometrial cells by reducing E₂ levels (Zhang *et al.*, 2013a). Moreover, Cao *et al.* (2017a) observed in cell

cycle analyses that curcumin increases the percentage of G1 phase cells in human endometriotic stromal cells, whereas the percentage of S phase cells and expression of VEGF is decreased. Hence, curcumin seems to reduce cell proliferation in human endometriotic stromal cells via down-regulation of the VEGF signalling pathway (Cao et al., 2017a). Recently, Chowdhury et al. (2019) detected a reduced secretion of different pro-angiogenic chemokines and pro-inflammatory cytokines from normal endometrial stromal cells and from eutopic endometrium of endometriosis subjects treated with curcumin. In contrast, the anti-inflammatory cytokines IL-10 and IL-12 were up-regulated by curcumin, particularly in endometrial stromal cells derived from endometriosis patients (Chowdhury et al., 2019). This indicates that the beneficial effects of curcumin on endometriosis may be mediated by an imbalance between pro- and anti-inflammatory factors.

In vivo, such beneficial curcumin effects have been reported by several groups in rat models of the disease (Yun-Fei et al., 2012; Kizilay et al., 2017; Jelodar and Azimifar, 2019). Yun-Fei et al. (2012) demonstrated that 150 mg/kg/day of intragastrically administered curcumin for three continuous weeks significantly reduces the volume and weight of endometriotic lesions. Because this was associated with a lower microvessel density within the lesions, they suggested that curcumin may inhibit the development of endometriosis by suppression of angiogenesis (Yun-Fei et al., 2012). This view is supported by another study of Zhang et al. (2011) demonstrating that curcumin doses of 50–150 mg/kg/day inhibit blood vessel formation in rat endometriotic lesions by down-regulating VEGF expression (Zhang et al., 2011).

In murine endometriotic lesions, curcumin has been shown to inhibit NF- κ B translocation as well as expression of MMP-2 and MMP-3 (Jana et al., 2012). Moreover, it stimulates apoptosis via the cytochrome-c-mediated mitochondrial pathway (Jana et al., 2012). In another study, 40 mg/kg/day curcumin encapsulated in poly(lactic-co-glycolic acid) nanoparticles suppressed oxidative stress parameters, angiogenic markers, matrix degrading molecules, oestrogen levels, endometriotic lesion size and microvessel density in the peritoneum of mice (Jana et al., 2014). In addition, Swarnakar and Paul (2009) reported that curcumin administered either pre- or post-induction of endometriosis inhibits the development of murine endometriotic lesions and reduces MMP-9 activity. Moreover, pretreatment with curcumin prevented lipid peroxidation and protein oxidation (Swarnakar and Paul, 2009). Hence, curcumin is a typical pleiotropic compound, which targets multiple molecular and cellular mechanisms in the pathogenesis of endometriosis (Arablou and Kolahdouz-Mohammadi, 2018).

Currently, the Vienna-based ENDOFLEX study is recruiting endometriosis patients for a randomised interventional clinical trial on placebo versus the dietary supplement flexofytol, with planned administration of two capsules containing 42 mg of curcumin twice a day for a duration of 4 months in the verum group. The study defines a possible change in the average pain score, from baseline to 4 months after the onset of treatment, as primary outcome, and changes in the number of days with pain, alleviation of dyspareunia, dysuria and dyschezia as well as changes in quality of life and sexual function as secondary outcomes (Perricos, 2020).

Puerarin

Puerarin is the main component of the root of *P. lobata*. This medicinal plant has a long history in China and possesses a wide range of

pharmacological properties (Zhang, 2019). The effects of puerarin on aromatase P450 (P450arom) expression in Ishikawa and RL95-2 endometrial carcinoma cell lines were assessed by Li et al. (2008). They demonstrated a significant decrease in P450arom expression at both mRNA and protein levels provoked by low-dose puerarin treatment in both cell lines (Li et al., 2008). In addition, puerarin suppressed E₂-induced proliferation of endometriotic stromal cells partly via down-regulation of a non-genomic membrane-initiated ERK pathway and inhibition of cyclin D1, COX-2 and Cyp19 gene expression (Cheng et al., 2012). Moreover, puerarin inhibited the invasion of endometriotic stromal cells and the vascularisation of ectopic endometrial tissues in the chicken chorioallantoic membrane model (Wang et al., 2011). This was associated with a reduced cellular accumulation of MMP-9, ICAM-1 and VEGF, whereas the expression of tissue inhibitors of metalloproteinases (TIMP)-1 increased after puerarin treatment (Wang et al., 2011). Ji et al. (2013) showed that puerarin suppresses the oestrogen-stimulated proliferation of endometriotic stromal cells partly through down-regulating the transcription of cyclin D1 and cdc25A (Ji et al., 2013).

In addition to these *in vitro* results, two studies also analysed the effect of puerarin in animal models of endometriosis. Chen et al. (2011b) orally administered the compound at three different doses, i.e. 60, 200 or 600 mg/kg/day, to rats, in which endometriotic lesions were successfully induced by transplanting autologous endometrial tissue to ectopic sites (Chen et al., 2011b). Even the lowest dose significantly reduced the weight of endometriotic tissue and serum oestrogen levels without adverse effects. Yu et al. (2015) demonstrated that puerarin inhibits the growth of rat endometriotic lesions due to the inhibition of P450arom and COX-2 expression. Puerarin further reduced the levels of E₂ and PGE₂ and blocked the positive feedback mechanism of E₂ synthesis by up-regulating the expression of 17 β -hydroxysteroid dehydrogenase-2 (17 β -hsd-2) and down-regulating the expression of 17 β -hydroxysteroid dehydrogenase-1 (17 β -hsd-1) (Yu et al., 2015).

Ginsenosides

Pharmacologic studies have shown that most of the biological activities of ginseng, the root of *Panax ginseng*, are derived from its main components, the ginsenosides (Dong-Soon and Seung-Yeol, 2013). A number of studies have reported that ginsenosides (e.g. ginsenoside-Rg3 and ginsenoside Rh2) and their metabolites (protopanaxadiol, PPD and protopanaxatriol, PPT) exhibit anti-tumour, anti-oxidant, immunomodulatory and anti-inflammatory activities (Zhang et al., 2018). Huang et al. (2020) recently exposed endometriotic stromal cells from patients with ovarian endometriosis to Rg3, which inhibited their proliferation in a time- and dose-dependent manner. In addition, Rg3 treatment significantly diminished the activity of NF- κ B, up-regulated the expression of caspase-3 and suppressed the expression of VEGF (Huang et al., 2020). In another study, Rg3 inhibited the development of endometriotic lesions in rats, which were induced by allotransplantation of endometrial tissue. The authors suggested that this effect is mediated by blockade of the VEGFR-2-activated PI3K/Akt/mTOR signalling pathway (Cao et al., 2017b). Qin et al. (2019) found that treatment with the ginsenoside Rf decreases the volume of endometriotic lesions and reduces the expression levels of VEGF and inflammation-related cytokines in rats. Of interest, they also showed that Rf inhibits

the expression of brain-derived neurotrophic factor, which could be involved in the alleviation of endometriosis-associated pain (Qin *et al.*, 2019). Zhang *et al.* (2018) observed that PPD, PPT, Rg3, Rh2 and esculetoside-A affect viability of, and induce autophagy in ectopic endometrial stromal cells. In addition, all compounds decreased the number and suppressed the growth of ectopic lesions in a murine endometriosis model (Zhang *et al.*, 2018).

Xanthohumol

Numerous studies have demonstrated the broad anti-cancer and anti-metastatic activities of xanthohumol, a flavonoid isolated from hops (*Humulus lupulus L.*), owing to its anti-proliferative, anti-inflammatory and anti-angiogenic properties (Jiang *et al.*, 2018). Accordingly, 100 μ M xanthohumol administered in the drinking water of mice with surgically induced endometriosis inhibited the development of endometriotic lesions independently of their localisation within the peritoneal cavity (Rudzitis-Auth *et al.*, 2012). Moreover, xanthohumol suppressed angiogenesis and cell proliferation within the lesions (Rudzitis-Auth *et al.*, 2012). Of interest, it was further found that treatment with xanthohumol did not affect the histomorphology, proliferation and vascularisation of the uterine horns and ovaries, suggesting that the treatment with this flavonoid may not induce side effects within the reproductive organs when treating endometriosis (Rudzitis-Auth *et al.*, 2012).

4-Hydroxybenzyl alcohol

4-HBA, one of the major active phenolic constituents of *Gastrodia elata* Blume, is an effective agent against several central and peripheral nervous disorders (Yu *et al.*, 2010). This phenolic compound exerts anti-oxidant, anti-inflammatory and anti-angiogenic actions on several experimental tumour models (Park *et al.*, 2010; Laschke *et al.*, 2013). Moreover, 4-HBA has been reported to inhibit the proliferation and migration of murine endothelial-like eEND2 cells, which is associated with a reduced expression of VEGF and MMP-9 (Laschke *et al.*, 2011). *In vivo*, 4-HBA suppressed the vascularisation and growth of developing endometriotic lesions in the dorsal skinfold chamber model (Laschke *et al.*, 2011).

Quercetin

Quercetin is ubiquitously present in fruits and vegetables, and is therefore one of the most common dietary flavonols in the Western diet. The anti-cancer effects of quercetin include its ability to reduce cell viability and increase apoptosis and autophagy through the modulation of PI3K/Akt/mTOR, Wnt/β-catenin and MAPK/ERK1/2 pathways (Reyes-Farias and Carrasco-Pozo, 2019). In rats with surgically induced endometriosis, quercetin inhibited the growth of endometriotic lesions through decreasing serum FSH and LH levels as well as reducing the local oestrogen content. Quercetin also decreased the expression of ER-α, ER-β and progesterone receptor in the hypothalamus, pituitary and endometrium, thereby inhibiting oestrogen and progesterone binding to their receptors (Cao *et al.*, 2014). In addition, Park *et al.* (2019) showed that quercetin inhibits cell proliferation and induces cell cycle arrest and apoptosis of the endocervical cell lines VK2/E6E7 and End1/E6E7. This was associated with the down-regulation of ERK1/2, P38 MAPK and AKT signalling molecules. Moreover, the

intraperitoneal administration of quercetin in a mouse model of endometriosis decreased the size of endometriotic lesions and exerted anti-proliferative and anti-inflammatory effects when compared to vehicle treatment in mice (Park *et al.*, 2019).

Apigenin

Apigenin is a natural bioactive flavonoid present in significant amounts within vegetables (parsley, celery, onions), fruits (oranges), herbs (chamomile, thyme, oregano, basil) and plant-based beverages (tea, beer, wine), which has been suggested to be suitable for the prevention and treatment of cancer, diabetes and Alzheimer's disease (Salehi *et al.*, 2019). Apigenin exerts anti-proliferative, anti-inflammatory and anti-angiogenic effects in cancer and metabolic diseases and, thus, has also been proposed as a potential therapeutic agent for endometriosis. In this context, apigenin has been shown to attenuate TNF-α-induced cell proliferation as well as COX-2 and PGE₂ synthesis via the inactivation of the NF-κB pathway in endometriotic stromal cells isolated from human endometriomas (Suou *et al.*, 2011). Moreover, Park *et al.* (2018) found that apigenin induces apoptosis in two human endometriotic cell lines through inhibition of phosphorylation of JNK and ERK1/2. Apigenin also reduced cell proliferation and induced cell cycle arrest in the two cell lines. However, the efficiency of the compound has not been evaluated so far in preclinical animal models of endometriosis (Park *et al.*, 2018).

Carnosic acid and rosmarinic acid

The phenolic diterpene, carnosic acid, and the polyphenol, rosmarinic acid, are the most abundant active compounds in rosemary leaves (*Rosmarinus officinalis*) and are thus responsible for the anti-oxidant activity of this plant (Moore *et al.*, 2016). Their anti-cancer (López-Jiménez *et al.*, 2013; Moore *et al.*, 2016) and anti-inflammatory effects (Yesil-Celiktas *et al.*, 2010; Petiwala and Johnson, 2015) have been demonstrated by several studies. Ferella *et al.* (2018) evaluated the effect of carnosic acid and rosmarinic acid on the development of experimental endometriosis. *In vitro*, carnosic acid and rosmarinic acid significantly inhibited cell proliferation in primary cultures and a human endometrial stromal cell line (T-HESC). Moreover, a strong anti-oxidant effect was observed after the exposure of T-HESC to rosmarinic acid, suggesting a possible role for reactive oxygen species (ROS) in the regulation of cell proliferation. *In vivo*, carnosic acid and rosmarinic acid significantly reduced the size of surgically induced endometriotic lesions in mice when compared to controls. Both compounds significantly suppressed cell proliferation, whereas rosmarinic acid additionally promoted apoptotic cell death within the lesions (Ferella *et al.*, 2018).

Wogonin and baicalein

Wogonin and baicalein are the main flavonoids isolated from the root of *Scutellaria baicalensis* (Li-Weber, 2009; Lee *et al.*, 2014). Their anti-tumour, anti-proliferative, anti-inflammatory and pro-apoptotic properties have already been demonstrated in different studies (Li-Weber, 2009; Wu *et al.*, 2016). Recent results provide evidence of an anti-proliferative effect of wogonin on human endometrial stromal cells, caused by the induction of cell cycle arrest at the G2/M phase (Ferella *et al.*, 2018). The inhibitory action of this flavonoid on the growth of

endometriotic lesions has been further demonstrated in a murine endometriosis model (Ferella et al., 2018). In addition, it was observed that baicalein significantly reduces the viability of human endometriotic stromal cells in culture, provokes G1 phase cell cycle and reduces the expression of the anti-apoptosis protein Bcl-2. The latter finding indicates that baicalein may induce apoptosis in human endometrial stromal cells (Jin et al., 2017).

Parthenolide

Parthenolide, a main active component of the traditional medical plant feverfew (*Tanacetum parthenium*), exhibits potent anti-cancer activities (Ghantous et al., 2013). In particular, parthenolide exerts anti-inflammatory, redox-modulating and epigenetic effects as well as selective cytotoxicity towards cancer stem and progenitor cells (Freund et al., 2020). In experimental studies, parthenolide inhibited the NF- κ B pathway and suppressed TNF- α -induced PGE₂ and DNA synthesis in endometrial stromal cells isolated from human endometriomas (Takai et al., 2013). In line with these findings, parthenolide further inhibited the formation of murine endometriotic lesions and the expression of pro-inflammatory-associated genes in a BALB/c mouse model of endometriosis (Takai et al., 2013).

Andrographolide

Andrographolide is the main active constituent present in *Andrographis paniculata*, a medicinal plant that has been widely used in complementary medicine, especially in Southeast Asia, China and India. Andrographolide has anti-thrombotic, anti-cancer and anti-inflammatory properties (Tan et al., 2017; Soo et al., 2019). Accordingly, andrographolide has been shown to inhibit cell cycle progression and cell proliferation through inhibition of the NF- κ B pathway and to inhibit COX-2 and tissue factor expression in primary cell cultures of endometriotic stromal cells. In addition, andrographolide-treated endometriotic lesions in rats exhibited a reduced size and expression of active NF- κ B subunits, nerve growth factor and COX-2. This was associated with a reduced pain behaviour of the treated animals, indicating that the compound is a potential anti-inflammatory and analgesic candidate for the treatment of endometriosis (Zheng et al., 2012).

Cannabinoids

Particularly in pain-associated pathologies, there is an increasing interest in the use of cannabis-based medicines. They contain cannabinoids derived from the cannabis plant, including delta-9-tetrahydrocannabinol (THC), the main psychoactive constituent of the *Cannabis sativa* plant, cannabidiol (CBD) or a combination of both (Boyaji et al., 2020). Two kinds of cannabinoid receptors are described: the cannabinoid 1 (CB1) and 2 (CB2) receptors. CB1 receptors are highly expressed in the uterus as well as in the spleen, heart, adrenal gland, ovaries and testes, among other tissues. The CB2 receptors are preferentially expressed in the immune system and intestine (Walker et al., 2019).

Some of the studies recently reviewed by Bouaziz et al. (2017) described endometriosis as an 'endocannabinoid deficiency' condition. It was reported that women with endometriosis have lower levels of CB1 receptors in their endometrial tissue (Bouaziz et al., 2017). Hence, the endocannabinoid system (ECS) could be important for the

establishment of pain associated with endometriosis, because reduced ECS function has been suggested to promote a more severe pain experience (Bouaziz et al., 2017).

The effects of subcutaneous administration of THC in a mouse model of surgically induced endometriosis were evaluated by Escudero-Lara et al. (2020). In this work, female mice, which develop mechanical hypersensitivity in the caudal abdomen, mild anxiety-like behaviour and substantial memory deficits associated with the presence of experimental endometriosis, were treated daily with THC (2 mg/kg). Interestingly, a moderate dose of the phytocannabinoid was effective in alleviating mechanical hypersensitivity and pain and restoring cognitive function without altering the anxiogenic phenotype. Strikingly, THC also inhibited the development of endometrial cysts (Escudero-Lara et al., 2020). These data highlight the interest of scheduled clinical trials designed to investigate possible benefits of THC for women with endometriosis. Regarding ongoing studies, the NIH-based database ClinicalTrials.gov only lists a currently recruiting phase 2 open label interventional clinical trial to study the effect of cannabinoid (THC/CBD 50%) on hyperalgesia in patients with deep endometriosis (Garcia Cinca, 2020), but no larger trials. It should be noted that targeting endocannabinoid modulation is probably even more than just treating pain, as it may also inhibit the growth of endometriotic tissue (Bouaziz et al., 2017). Hence, this type of phytotherapy has great potential for clinical use. Nonetheless, more investigation is needed to evaluate its efficiency and potential therapeutic side effects during chronic use, especially on fertility and pregnancy outcomes (Bouaziz et al., 2017).

Summary

As summarised in Table II, several specific plant-derived bioactive compounds have been shown to exert beneficial effects on endometriotic cells or lesions. In contrast to conventional drugs, they typically do not selectively inhibit one molecular target, but exhibit a pleiotropic action profile, which may simultaneously affect multiple signalling pathways. While this can markedly increase their efficacy in the treatment of endometriosis, it also complicates the identification of key mechanisms that mediate their therapeutic effects. Another important issue is the fact that most plant-derived bioactive compounds exhibit a low *in vivo* stability and bioavailability, which makes their efficient use in endometriosis patients difficult (Wieser et al., 2007). To overcome this problem, it will be necessary to identify analogues or to develop novel derivatives or prodrugs with improved pharmacokinetic properties. Moreover, sophisticated carrier systems are required, to selectively accumulate in endometriotic tissue, leading to high local compound concentrations without inducing systemic side effects. While challenging, these goals appear realistic considering the rapid progress in modern drug design and nanotechnology. Accordingly, the first promising steps towards this direction have already been taken for resveratrol, EGCg, quercetin, baicalein and curcumin (Davatgaran-Taghipour et al., 2017; Shi et al., 2018; Intagliata et al., 2019; Vinayak and Maura, 2019). For instance, demethoxycurcumin, a naturally occurring curcumin analogue, which lacks the methoxy group on the benzene ring of the parent structure, exhibits a much higher chemical stability at physiological pH (Hatamipour et al., 2019). Hence, this curcuminoid has already been suggested for the treatment of different diseases, such as cancer, malaria, Parkinson's disease, rheumatoid arthritis, inflammatory bowel

Table II Plant-derived bioactive compounds evaluated in experimental models of endometriosis.

| Bioactive compound | Model/dosage | Action/mechanism | Reference |
|--------------------|--|--|------------------------------------|
| Resveratrol | Primary human endometrial stromal cells | ↓ Invasiveness | Bruner-Tran <i>et al.</i> (2011) |
| | Primary human endometrial epithelial cells | ↓ Cell proliferation | Ricci <i>et al.</i> (2013) |
| | Primary human endometriotic stromal cells | ↑ Apoptosis ↑ TRAIL-induced apoptosis ↓ Survivin | Taguchi <i>et al.</i> (2016) |
| | Primary human endometriotic stromal cells | ↓ TNF- α -induced IL-8 release | Taguchi <i>et al.</i> (2014) |
| | Ishikawa epithelial endometrial cell line | ↓ IGF-I and HGF levels | Arablou <i>et al.</i> (2019) |
| | Heterologous model of endometriosis in nude mice (6 mg/mouse by gavage) | ↓ Number of lesions ↓ Endometriotic lesion size ↑ Apoptosis | Bruner-Tran <i>et al.</i> (2011) |
| | Xenograft implants of human endometrial tissue in immunodeficient RAG-2-g(c) mice (6, 30 or 60 mg/day s.c.) | ↓ Expression of ESRI (with 60 mg/day) ↓ Proliferative activity (with 60 mg/day) | Amaya <i>et al.</i> (2014). |
| | Balb/c mice with induced endometriosis (10 or 25 mg/kg/day i.p.) | ↓ Cell proliferation ↑ Apoptosis ↓ Number of lesions (with 25 mg/kg/day) | Ricci <i>et al.</i> (2013) |
| | Balb/c mice with induced endometriosis (40 mg/kg/day by gavage) | ↓ Endometriotic lesion size ↓ Cell proliferation ↓ Angiogenesis | Rudzitis-Auth <i>et al.</i> (2013) |
| | Sprague-Dawley rats with induced endometriosis (10 mg/kg/day i.m.) | ↓ Endometriotic lesion size ↓ VEGF levels ↓ MCP-1 levels | Ergenoglu <i>et al.</i> (2013) |
| | Wistar rats with induced endometriosis (60 mg/kg/day by gavage) | ↓ Histopathological scores of endometriotic lesions ↓ VEGF levels ↓ MCP-1 levels | Cenksoy <i>et al.</i> (2015) |
| | Wistar rats with induced endometriosis (1 or 10 mg/kg/day i.p.) | ↓ Endometriotic lesion size ↓ Cell proliferation ↑ Superoxide dismutase and glutathione ↑ Malonyl dialdehyde and catalase | Yavuz <i>et al.</i> (2014) |
| EGCG | Sprague-Dawley rats with induced endometriosis (1 or 30 mg/kg/day i.m.) | ↓ Endometriotic lesion size ↓ IL-6, IL-8 and TNF- α ↓ VEGF levels ↓ MMP-2 and MMP-9 expression | Bayoglu Tekin <i>et al.</i> (2015) |
| | Hamster endometrial glandular cells and endometrial stromal cells | ↓ Cell proliferation ↓ VEGF levels | Laschke <i>et al.</i> (2008) |
| | Primary human endometrial epithelial cells | ↓ Cell proliferation ↑ Apoptosis | Ricci <i>et al.</i> (2013) |
| | Primary human endometrial and endometriotic stromal cells | ↓ Cell proliferation ↓ Invasion ↓ Migration ↓ Fibrotic markers ↓ TGF- β 1-stimulated activation of MAPK and Smad | Matsuzaki and Darcha (2014) |
| | Endometrial fragments transplanted into the dorsal skinfold chamber of female Syrian golden hamsters (65 mg/kg/day i.p.) | ↓ Endometriotic lesion size ↓ Angiogenesis | Laschke <i>et al.</i> , (2008) |
| | Heterologous immunocompromised mouse model of experimental endometriosis (50 mg/kg/day i.p.) | ↓ Endometriotic lesion size ↑ Apoptosis ↓ Angiogenesis ↓ VEGF levels ↓ VEGFR-2 expression | Xu <i>et al.</i> (2009) |
| | Balb/c mice with induced endometriosis (20 or 100 mg/kg/day by gavage) | ↓ Endometriotic lesion size ↓ Cell proliferation ↑ Apoptosis | Ricci <i>et al.</i> (2013) |
| | Heterologous model of endometriosis in nude mice (50 mg/kg/day i.p.) | ↓ Fibrosis | Matsuzaki and Darcha (2014) |

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Table II Continued

| Bioactive compound | Model/dosage | Action/mechanism | Reference |
|--------------------|---|--|-----------------------------|
| Curcumin | Human endometriotic stromal cells from ovarian endometrioma | ↓ ICAM-1 and VCAM-1 expression ↓ IL-6, IL-8 and MCP-1 secretion ↓ Activation of NF-κB-induced by TNF- α | Kim et al. (2012) |
| | Human endometriotic cells and normal endometrial cells | ↓ Cell proliferation ↓ E ₂ levels | Zhang et al. (2013a) |
| | Human endometriotic stromal cells from endometriosis patients | ↓ Cell proliferation ↑ Percentage of G1 phase cells ↓ Percentages of S phase cells ↓ VEGF levels | Cao et al. (2017a) |
| | Human endometriotic stromal cells from endometriosis patients | ↓ Pro-inflammatory and pro-angiogenic chemo-kine and cytokine secretion ↑ IL-10 and IL-12 secretion | Chowdhury et al. (2019) |
| | Surgically induced endometriosis in rats (150 mg/kg/day by gavage) | ↓ Endometriotic lesion size ↓ Microvessel density | Yun-Fei et al. (2012) |
| | Wistar rats with induced endometriosis (50, 100 or 150 mg/kg/day by gavage) | ↓ Endometriotic lesion size ↓ Angiogenesis ↓ VEGF levels | Zhang et al. (2011) |
| | Wistar rats with induced endometriosis (100 mg/kg/day by gavage) | ↓ Endometriotic lesion size ↓ Cell proliferation | Kizilay et al. (2017) |
| | Sprague-Dawley rats with induced endometriosis (48 mg/kg/day i.p.) | Suppression of the growth of endometriotic lesions | Jelodar and Azimifar (2019) |
| | Balb/c mice with induced endometriosis (protective effect: 16, 24 or 48 mg/kg i.p. 30 min prior the induction of endometriosis and once daily for the next 3 days; therapeutic effect: 48 mg/kg/day i.p. for 10 days) | ↓ Endometriotic lesion size ↑ Apoptosis ↓ NF-κB translocation ↓ MMP-3 expression | Jana et al. (2012) |
| | Swiss Albino mice with induced endometriosis (40 mg/kg/day encapsulated in PLGA nanoparticles) | ↓ Endometriotic lesion size ↓ Oxidative stress parameters ↓ Angiogenesis ↓ Oestrogen levels ↓ Lipid peroxidation ↓ Protein oxidation ↓ MMP-9 activity | Jana et al. (2014) |
| Puerarin | Balb/c mice with induced endometriosis (protective effect: 16, 32 or 48 mg/kg i.p. 30 min prior the induction of endometriosis and once daily for the next 3 days; therapeutic effect: 48 mg/kg/day i.p. for 10 days) | ↓ Endometriotic lesion size ↓ Oxidative stress parameters ↓ Angiogenesis ↓ Oestrogen levels ↓ Lipid peroxidation ↓ Protein oxidation ↓ MMP-9 activity | Swarnakar and Paul (2009) |
| | Ishikawa and RL95 endometrial cell lines | ↓ P450Arom expression and activity | Li et al. (2008) |
| | Human endometriotic stromal cells from ovarian endometrioma | ↓ Cell proliferation | Cheng et al. (2012) |
| | Human endometriotic stromal cells from ovarian endometrioma | ↓ Cyclin D1, Cox-2, Cyp19 expression ↓ Invasion ↓ Vascularisation ↓ MMP-9 | Wang et al. (2011) |
| | Human endometriotic stromal cells from ovarian endometrioma | ↓ ICAM ↓ VEGF ↓ TIMP-1 | Ji et al. (2013) |
| | Sprague-Dawley rats with induced endometriosis (60, 200 or 600 mg/kg/day by gavage) | ↓ Cell proliferation ↓ Cyclin D1, cdc25A ↓ Endometriotic lesion size ↓ P450Arom expression | Chen et al. (2011b) |
| | Sprague-Dawley rats with induced endometriosis (5, 20 or 80 mg/kg/day i.p.) | ↓ E ₂ levels ↓ Endometriotic lesion size ↓ P450Arom ↓ COX-2 ↓ E ₂ levels ↓ PGE ₂ ↑ 17 β -hsd-2 ↓ 17 β -hsd-1 | Yu et al. (2015) |

Table II Continued

| Bioactive compound | Model/dosage | Action/mechanism | Reference |
|--|--|--|------------------------------------|
| Ginsenosides | Human endometriotic stromal cells from ovarian endometrioma Rg3 | ↓ Cell proliferation ↓ NF-κB activity ↓ VEGF ↑ Caspase-3 | Huang <i>et al.</i> (2020) |
| | Human ectopic endometrial cells PPD, PPT, Rg3, Rh2 or EsA | ↓ Cell proliferation ↑ Autophagy | Zhang <i>et al.</i> (2018) |
| | Sprague-Dawley rats with induced endometriosis (5 or 10 mg/kg/day of Rg3) | ↓ Endometriotic lesion size ↓ VEGFR-2 mediated PI3K/Akt/mTOR signaling pathway | Cao <i>et al.</i> (2017b) |
| | Wistar rats with induced endometriosis (1, 2 or 4 mg/kg/day i.p. of Rf) | ↓ Endometriotic lesion size ↓ VEGF ↓ iNOS ↓ IL-6 ↓ IL-1 β ↓ TNF- α ↓ BDNF | Qin <i>et al.</i> (2019) |
| | Balb/c mice with induced endometriosis (45, 30 or 15 mg/kg of PPD, PPT, Rg3, Rh2 or EsA on day 4 and day 10, i.p.) | PPD, PPT, Rg3, Rh2 or EsA ↓ Endometriotic lesion size ↓ Number of lesions | Zhang <i>et al.</i> (2018) |
| | Balb/c mice with induced endometriosis (100 μ M xanthohumol via drinking water) | ↓ Endometriotic lesion size ↓ PI3 kinase ↓ Angiogenesis ↓ Cell proliferation | Rudzitis-Auth <i>et al.</i> (2012) |
| 4-HBA | Murine endothelial-like eEND2 cells | ↓ Cell proliferation ↓ MMP-2 and VEGF ↓ Cell migration | Laschke <i>et al.</i> (2011) |
| | Endometrial fragments transplanted into the dorsal skinfold chamber of C57BL/6 mice (100 mg/kg/day i.p.) | ↓ Endometriotic lesion size ↓ Angiogenesis | Laschke <i>et al.</i> (2011) |
| Quercetin | Sprague-Dawley rats with induced endometriosis (60, 150 or 375 mg/kg/day i.p.) | ↓ Serum FSH and LH levels ↓ Serum oestrogen content ↓ ER α , ER β and PR in hypothalamus, pituitary and endometrium | Cao <i>et al.</i> (2014) |
| | Endocervical cell lines VK2/E6E7 and End1/E6E7 | ↓ Cell proliferation ↓ ERK1/2, P38 MAPK and AKT ↑ Cell cycle arrest | Park <i>et al.</i> (2019) |
| | C57BL/6 mice with induced endometriosis (35 mg/kg/day i.p.) | ↓ Endometriotic lesion size ↓ Cell proliferation ↓ Inflammation | Park <i>et al.</i> (2019) |
| | VK2/E6E7 and End1/E6E7 endometriosis cell lines | ↓ Cell proliferation ↑ Apoptosis ↑ Cell cycle arrest | Park <i>et al.</i> (2018) |
| Apigenin | Human endometriotic stromal cells from ovarian endometrioma | ↓ Cell proliferation ↓ I κ B phosphorylation ↓ COX-2 ↓ PGE ₂ | Suou <i>et al.</i> (2011) |
| | Primary endometrial stromal cells from patients with endometriosis | ↓ Cell proliferation ↓ Oxidation | Ferella <i>et al.</i> (2018) |
| Carnosic acid and rosmarinic acid | Human endometrial stromal cell line (T-HESC) | ↓ Endometriotic lesion size | Ferella <i>et al.</i> (2018) |
| | Balb/c mice (CA 2mg/kg/day or 20mg/kg/day i.p.) | ↓ Cell proliferation | Ferella <i>et al.</i> (2018) |
| | (RA 1mg/kg/day or 3mg/kg/day i.p.) | ↑ Apoptosis | |

Table II Continued

| Bioactive compound | Model/dosage | Action/mechanism | Reference |
|-------------------------------------|---|--|--|
| Wogonin | Primary endometrial stromal cells from patients with endometriosis Human endometrial stromal cell line (T-HESC) Balb/c mice (20mg/kg/day by gavage) | ↓ Cell proliferation ↑ Cell cycle arrest ↓ Endometriotic lesion size ↓ Cell proliferation ↑ Apoptosis | Ferella et al. (2018) Ferella et al. (2018) |
| Baicalein | Human endometriotic stromal cells from ovarian endometrioma | ↓ Cell proliferation G1 phase cell cycle arrest ↓ Bcl-2 | Jin et al. (2017) |
| Parthenolide | Human endometriotic stromal cells from ovarian endometrioma Balb/c mice (10 mg/kg/thrice weekly) | ↓ TNF- α -induced IL-8 release ↓ COX-2 ↓ PGE ₂ ↓ Cell proliferation ↓ I κ B phosphorylation ↓ Number of lesions ↓ Endometriotic lesion size ↓ Cell proliferation ↓ VEGF ↓ IL-6 ↓ MCP-1 ↓ LIF | Takai et al. (2013) Takai et al. (2013) |
| Andrographolide | Primary cell cultures of endometriotic stromal cells Sprague–Dawley rats with induced endometriosis (50 or 187.5 mg/kg by gavage) | ↓ Cell cycle progression ↓ Cell proliferation ↓ NF- κ B activation ↓ COX-2 ↓ Tissue Factor ↓ Endometriotic lesion size ↓ Endometriosis-pain ↓ NF- κ B activation ↓ NGF ↓ COX-2 | Zheng et al. (2012) Zheng et al. (2012) |
| Delta-9-tetrahydrocannabinol | C57Bl/6J mice (2 mg/kg/day) | ↓ Endometriotic lesion size ↓ Endometriosis-pain ↓ Hyperinnervation | Escudero-Lara et al. (2020) |

BDNF, brain derived neurotrophic factor; 17 β -hsd, 17 β -hydroxysteroid dehydrogenase; CA, carnosic acid; E2, oestrogen; EsA, esculentoside A; ESRI, oestrogen receptor I; HGF, hepatocyte growth factor; IGF-I, insulin-like growth factor-I; IL-1 β , interleukin-1 β ; IL-8, interleukin-8; iNOS, inducible nitric oxide synthase; LIF, leukemia inhibitory factor; MCP-1, monocyte chemoattractant protein-1; NGF, nerve growth factor; P450Arom, aromatase P450; PLGA, poly(lactic-co-glycolic acid); PPD, 20(S)-protopanaxadiol; PPT, 20(S)-protopanaxatriol; PR, progesterone receptor; RA, rosmarinic acid; Rf, ginsenoside Rf; Rg3, ginsenoside Rg3; Rh2, ginsenoside Rh2; T-HESC, human endometrial stromal cell line; TIMP, tissue inhibitor of MMP; TRAIL, tumour necrosis factor-related apoptosis-inducing ligand; TRPV-1, transient receptor potential cation channel subfamily V member 1; VEGF, vascular endothelial growth factor; VEGF-R, vascular endothelial growth factor receptor.

disease and hypertension (Hatamipour et al., 2019). In addition, the oral bioavailability of curcumin may be improved by means of solid dispersions, nanoparticles, polymeric micelles, nanosuspensions and lipid-based nanocarriers (Ma et al., 2019). Moreover, distinct curcumin formulations with excellent human bioavailability and pharmacokinetics data are now available. For instance, NovaSol[®], CurcuWin[®] and LongVida[®] have been reported to exhibit an over 100-fold higher bioavailability relative to unformulated curcumin (Jamwal, 2018).

Finally, specific plant-derived bioactive compounds have already entered the important stage of clinical evaluation. However, the current study situation is far from sufficient to justify the inclusion of such compounds in modern multimodal treatment concepts for endometriosis.

This can only be achieved based on future randomised controlled multi-centre trials including large patient cohorts.

Chinese Herbal Medicine

As one of the major contemporary alternative medicines, traditional Chinese herbal medicine (CHM) is not only of great significance in China, but meanwhile also attracts the academic attention in the world of Western medicine. CHM traditionally uses polypharmacological interactions of herbal formulae with multiple ingredients to treat complex diseases. The idea is that each individual herb works synergistically to treat the disease process (Burks-Wicks et al., 2005). In contrast to

Table III Ingredients of Chinese herbal medicine formulae.

| Formula | Ingredients | Reference |
|---------|---|----------------------|
| PCJNF | Ramulus Cinnamomi, Sanguis Draconis, Faeces Tropopterori, Pollen Typhae, Chinese Eaglewood, <i>Whitmania pigra</i> Whitman, <i>Liriope spicata</i> , <i>Glycyrrhiza uralensis</i> Fisch | Liang et al. (2017) |
| XLF | Fried turtle shells, sliced <i>Cornus Cervi</i> , leech, <i>Eupolyphaga</i> seu <i>Steleophaga</i> , <i>Vaccaria segetalis</i> , <i>Rhizoma curcumae</i> , steamed buns, <i>Salvia chinensis</i> , <i>Smilax china</i> , <i>Radix codonopsis</i> and <i>Rhizoma zingiberis</i> Preparatum | Zhou et al. (2019) |
| WXT | <i>Typha angustifolia</i> L., <i>Tropopterus xanthipes</i> Milne-Edward, <i>Draconis sanguis</i> , <i>Leguminosae</i> , <i>Radix Dipsaci Asperoidis</i> , <i>Commiphora myrrha</i> , <i>Corydalis yanhusuo</i> W.T.Wang, <i>Fructus Meliae</i> Toosendan | Zhang et al. (2013b) |
| GZFLC | Ramulus Cinnamomi, <i>Poria</i> , <i>Semen Persicae</i> , <i>Radix Paeoniae</i> , <i>Rubra</i> or <i>Radix Paeoniae Alba</i> , <i>Cortex Moutan</i> | Zhou et al. (2018) |
| GZFLW | | |
| SJZTC | <i>Drac(h)onis sanguis</i> , <i>Panax pseudoginseng</i> (saponin), <i>Thunberg fritillary bulb</i> (Verticinone), <i>Coix</i> seed | Zou et al. (2013) |
| XCHT | <i>Bupleurum chinense</i> , <i>Scutellaria baicalensis</i> , ginseng root, <i>Pinellia</i> , licorice, ginger, jujube | Jiao et al. (2013) |
| HYXZ | <i>Salvia miltiorrhiza</i> radix, <i>Morinda officinalis</i> radix, <i>Panax notoginseng</i> radix, <i>Semen coicis</i> , <i>Fritillariae thunberg</i> bulbus, <i>Spica prunellae</i> , <i>Polygoni aviculare</i> herba, <i>Panta rhei</i> radix, <i>Alternaria dianthi</i> herba, <i>Corydalis</i> sp. rhizome, <i>Hirudo</i> sp., <i>Typhae</i> sp. Pollen, <i>Draconis sanguis</i> | Chen and Gong (2017) |
| SZD | <i>Foeniculi Fructus</i> , <i>Zingiberis Rhizoma</i> , <i>Corydalis Rhizoma</i> , <i>Myrrha</i> , <i>Rhizoma Chuanxiong</i> , <i>Angelicae Sinensis</i> Radix, <i>Radix Paeoniae Rubra</i> , <i>Cortex Cinnamomi</i> , <i>Typhae Pollen</i> , <i>Tropopteri Feces</i> | Zhu et al. (2018) |
| JFS | Ligustrazine, ferulic acid and tetrahydropalmatine | Chen et al. (2018b) |
| TSS | <i>Angelicae radix</i> , <i>Hoelen</i> , <i>Cnidii rhizoma</i> , <i>Alismatis rhizoma</i> , <i>Paeoniae Radix</i> , <i>Atractylodis lancea</i> rhizome | Nagira et al. (2019) |
| BSHXR | <i>Astragali radix</i> , <i>Angelicae sinensis</i> radix, <i>Ligustici Chuanxiong</i> Rhizoma, <i>Cuscutae semen</i> , <i>Taxilli Herba</i> , <i>Dipsaci Radix</i> | Ding et al. (2018) |
| DEFK | <i>Salvia</i> , <i>Rhizoma Curcumae</i> , <i>Rhizoma Sparganii</i> , <i>bupleurumroot</i> , <i>angelica</i> , <i>liquorice</i> , <i>Rhizoma Corydalis</i> , <i>Radix Paeoniae Rubra</i> , <i>Rhizoma Cyperi</i> | Li et al. (2017) |
| XFZY | <i>Persicae Semen</i> , <i>Flos Carthami Tinctorii</i> , <i>Radix Angelicae Sinensis</i> , <i>Radix Rehmanniae Glutinosae</i> , <i>Rhizoma Ligustici Chuanxiong</i> , <i>Radix Rubrus Paeoniae Lactiflorae</i> , <i>Achyranthis Bidentatae Radix</i> , <i>Platycodi Radix</i> , <i>Radix Bupleuri</i> , <i>Fructus Aurantii</i> , <i>Radix Glycyrrhizae</i> | Zhang et al. (2012) |

BSHXR, Bushen Huoxue recipe; DEFK, Danefukang; GZFLC, Gui-Zhi-Fu-Ling Capsule; GZFLW, Gui-Zhi-Fu-Ling-Wan; HYXZ, Hua Yu Xiao Zheng; JFS, Jiawei-Foshou-San; PCJNF, Ping-Chong-Jiang-Ni; SJZTC, Sanjie Zhentong capsule; SZD, Shaofu Zhuyu Decoction; TSS, tokishakuyakusan; WXT, Wenshen Xiaozheng Tang; XCHT, Xiaochaihu Tang; XFZY, Xuefu-Zhuyu capsule; XLF, Xiao Liu Fang.

Western medicine, which focuses on fighting the disease directly, treatment with CHM emphasises the prescription for individual patients' syndrome patterns and holistically attempts to adjust the body's physiology. For example, a part of endometriosis-associated symptoms are summarised in CHM as 'blood-stasis syndrome' (Wieser et al., 2007; Guo et al., 2010), which is reflected by the anti-platelet, anti-thrombotic and anti-coagulation activity of some CHM formulas (Table III). Chinese herbs are generally taken as pills or decoctions (teas) while the design of the formula and the dosage varies. By now, several herbal products are also available for the treatment of endometriosis and infertility.

Formulations for endometriosis treatment

There are several representative classic Chinese herbal prescriptions containing *Cinnamomi ramulus*, an ancient herbal medicine used for thousands of years in China with a broad spectrum of pharmacological activities (Liu et al., 2020). One of them is the formula Ping-Chong-Jiang-Ni (PCJNF, Table III) which is clinically effective on dysmenorrheal relief and reduction of CA-125 and prolactin serum levels in endometriosis patients (Liang et al., 2017). *In vitro*, PCJNF suppresses proliferation, migration and invasion while increasing apoptosis in ectopic endometrial stromal cells, which is mediated by JNK signalling (Liang et al., 2017).

A preliminary study reported that the Xiao Liu Fang (XLF, Table III) formula is effective in the treatment of uterine fibroids (Wen, 2007; Liu et al., 2013). *In vitro*, XLF extract has an effect similar to that of gestrinone on ectopic and eutopic endometrial stromal cells of endometriosis patients and endometrial tissue of healthy women, as it inhibits endometrial cell attachment and angiogenesis by reducing the cellular expression levels of ICAM-1, COX-2, MMP-9 and VEGF mRNA (Zhou et al., 2019).

Zhang et al. (2016) evaluated the effect of Wenshen Xiaozheng Tang (WXT, Table III) on primary ectopic endometriotic stromal cells. WXT inhibited the proliferation and migration of the cells and induced their apoptotic cell death by the activation of caspases, by reducing NF- κ B activation, thus lowering the expression of NF- κ B-regulated gene products such as cellular inhibitor of apoptosis protein (c-IAP)1, c-IAP2, X-linked inhibitor of apoptosis protein, survivin, myeloid leukemia cell differentiation protein (Mcl)-1, COX-2 and MMP-9 (Zhang et al., 2016). *In vivo*, WXT significantly decreased the mean size of endometriotic lesions as well as the peritoneal fluid and serum levels of TNF- α and IL-1 β in a rat model of endometriosis. In addition, it down-regulated the expression of COX-2, MMP-9, plasminogen activator inhibitor and ICAM-1 and up-regulated the expression of TIMP1 within the lesions, indicating that WXT inhibits the production of pro-inflammatory cytokines and regulates the expression of invasion-related genes in endometriosis (Zhang et al., 2013b).

Gui-Zhi-Fu-Ling-Wan (GZFLW, **Table III**) is one of the most frequently prescribed CHM that has been used in the therapy of women with endometriosis to relieve inflammation-related symptoms (Fang et al., 2012). Gui-Zhi-Fu-Ling capsules (GZFLC) originate from the classic Chinese medicinal formula GZFLW, which is more convenient to be taken and easily accepted by the majority of patients (Zhou et al., 2018). A few studies have been conducted to investigate the effects of GZFLC in relation to the metabolic changes in pelvic endometriosis. Ji et al. (2011) evaluated autologous endometrial explants in a rat model of surgically induced endometriosis and observed that the volume of developing endometriotic lesions was significantly reduced in the GZFLC treatment group when compared to controls. This was associated with ICAM-1 and MCP-1 down-regulation within the lesions. Moreover, the numbers of CD4⁺ T lymphocytes and the activity of natural killer cells were higher than in the control group (Ji et al., 2011). In the same model, TUNEL assays showed that different doses of GZFLC induce apoptosis in endometriotic cells. GZFLC further inhibited the mRNA levels of the anti-apoptotic Bcl-2 gene and increased the mRNA levels of the pro-apoptotic factor Bcl-2-associated X protein. These results indicate that GZFLC inhibits cell proliferation and induces apoptosis of endometriotic cells through the mitochondrial apoptotic pathway (Hu et al., 2014). Furthermore, proton nuclear magnetic resonance spectroscopy-based targeted metabolite profiling demonstrated that GZFLC significantly affects the expression levels of transforming growth factor (TGF)- β 1, glucose transporter (GLUT)-4 and VEGF, indicating that GZFLC also affects endometriosis by influencing glycolysis and gluconeogenesis (Zhou et al., 2018).

Sanjie Zhentong capsule (SJZTC, **Table III**) exhibits immunoregulatory and anti-inflammatory activity and relieves pain. In a rat model of surgically induced endometriosis, SJZTC reduced the size of endometrial explants, most likely by inhibiting the expression of VEGF and TNF- α (Zou et al., 2013).

Another classic Chinese herbal formulation with anti-inflammatory and immunoregulatory effects is Xiaochaihu Tang (XCHT, **Table III**) (Jiao et al., 2013; Zhao et al., 2017). XCHT exerts a therapeutic effect on endometriosis in rats through the reduction of MMP-2 and MMP-9 activity in ectopic endometrial tissue (Jiao et al., 2013).

In clinical practice, it has been observed that Hua Yu Xiao Zheng (HYXZ, **Table III**) decoction alleviates endometriosis-associated symptoms, including severe dysmenorrhea, dyspareunia, menstrual irregularities and infertility (Chen and Gong, 2017). *In vivo*, this formula decreases the size of endometriotic lesions in rats and affects angiogenesis through reduction of VEGF and angiopoietin-2 expression (Chen and Gong, 2017).

In Taiwan, one of the top five commonly used herbal formulas for the treatment of endometriosis is the Shaofu Zhuyu Decoction (SZD, **Table III**), a classic traditional Chinese medicine for dysmenorrhea, which has also been widely used in clinical practice to relieve symptoms of endometriosis (Tsai et al., 2017). In a rat endometriosis model, SZD significantly reduced the size of ectopic lesions, inhibited cell proliferation, increased apoptosis and reduced microvessel density and hypoxia-inducible factor expression within the lesions (Zhu et al., 2018).

Jiawei-Foshou-San (JFS, **Table III**) formula has been demonstrated to diminish the growth of endometriotic lesions, reduce oestrogen levels and suppress inflammation and angiogenesis in rats (Tang et al., 2014). Chen et al. (2018b) developed an autologous transplantation model

for the induction of endometriosis in rats to evaluate the *in vivo* effect of JFS. They observed that JFS significantly suppresses the growth and reduces the volume of ectopic endometrium. Moreover, JFS inhibited the gene expression of MMP-2, MMP-9 and up-regulated the mRNA levels of TIMP-1 (Chen et al., 2018b).

CHM has already been used to treat patients with endometriosis with favourable effects on disease-related symptoms and fertility (Ried and Stuart, 2011; Bina et al., 2019). In fact, a Cochrane study evaluating randomised controlled trials reported that post-surgical administration of CHM may be comparably effective as gestrinone but with fewer side effects (Flower et al., 2012). Moreover, treatment with CHM may be even better than danazol (Flower et al., 2012). In addition, it was suggested that CHM results in higher pregnancy rates when compared to Western medical fertility drug therapy (Ried, 2015).

In a randomised multi-centre trial involving 208 patients, CHM was compared with Western medicine for the treatment of endometriosis (Zhao et al., 2013). The CHM-treated groups received three types of formulations based on their particular syndrome differentiation. Patients in the Western medicine group were treated with GnRHa or gestrinone. Treatment was started by administration of the drugs to the patients during days 1–5 of their first menstruation in both groups. The patients with stages I and II of endometriosis were treated for 3 months, while the patients with stages III and IV were treated for 6 months (ASRM, 1997). Although, the incidence and timing of recurrence of the disease were not significantly different between the two groups, the first pregnancy in the CHM group was significantly earlier than the first pregnancy in the Western medicine group. Moreover, CHM had some advantages, such as low incidence and mild symptoms of adverse reactions as well as good patient compliance (Zhao et al., 2013).

In a large study carried out in Taiwan, the utilisation of CHM among women with endometriosis was analysed. For this purpose, a randomly sampled cohort of 1 000 000 beneficiaries recruited from the Taiwan National Health Insurance Research Database was evaluated and 14 080 study subjects were included in the study. Among them, 12 788 (90.8%) endometriosis patients used CHM at least once during the 11-year study period. Patients suffering from the symptoms of endometriosis were more likely to choose CHM treatment than those with no symptoms. GZFLW was the most frequently prescribed CHM among users. The potential effects of these Chinese herbs when used to treat endometriosis are related to their anti-inflammatory, anti-proliferative and pain-alleviating properties (Fang et al., 2012). Despite the very large number of subjects involved in this study, the non-randomised and uncontrolled methodology of the studies prevents definitive conclusions and emphasises the need for well-conducted, double-blind, randomised, placebo-controlled studies to further evaluate the efficacy of GZFLW and other medicinal herb formulations on women with endometriosis (Guo et al., 2010). Due to the limitations of this work, the authors suggested that the results could have been confounded by a placebo effect (Fang et al., 2012).

More recently, Tsai et al. (2017) conducted an additional study with the objective to identify a CHM network for endometriosis by analysing the same nationwide CHM prescription database in Taiwan. From 2008 to 2013, a total of 33 235 endometriosis patients were identified and 31 650 patients used CHM at least once for any reason. Among all herbal formulae, GZFLW was used most frequently (28.1% of all

prescriptions) and *Cyperus rotundus* was the most commonly used single herb (18.8% of all prescriptions) (Tsai *et al.*, 2017).

An additional Taiwanese population-based retrospective cohort study evaluated the relationship between the use of CHM and subsequent surgery among patients with endometriosis (Su *et al.*, 2014). A total of 8283 CHM users were identified among the 22 488 endometriosis patients found in the National Health Insurance reimbursement database between 2000 and 2010, while a control group consisted of 8283 women with endometriosis who were non-users of CHM. The treated group included subjects, who had received an orally administered CHM treatment for more than 2 consecutive weeks. This study found that in endometriosis patients, the incidence rate and risk of surgery were both lower among CHM users than among non-users. Moreover, surgery was performed earlier in the group of non-users than in the group of CHM users. Hence, CHM appears to provide an alternative treatment option that reduces the incidence of surgery and, thus, could have important health economic implications (Su *et al.*, 2014).

Another clinical study performed by Ding and Lian (2015) evaluated a total of 80 patients treated with hormone or traditional CHM. Patients belonging to the hormone therapy group took 12.5 mg mifepristone orally every day. The therapy started from the 1st day of their menstrual cycle and lasted for 6 months. In the traditional Chinese medicine group, two kinds of herbal mixtures were given to the patients in different periods of their menstrual cycle. The first one was applied before ovulation, while the other was given after ovulation. After water decoction, each herb was taken separately in the morning and evening (Ding and Lian, 2015). After treatment, serum oestradiol, FSH and CA-125 levels decreased. In addition, CHM has demonstrated to be safe. No significant changes on serum levels of glutamic-pyruvic transaminase and creatinine were found. At the same time, there were no severe adverse events during the follow-up treatment and the pregnancy rate was similar in both groups. There were no cases of ectopic pregnancy and no significant differences in spontaneous abortion rate were observed. Despite the limitations of this approach (small sample size and the fact that surgery was not taken into account), the study provided preliminary data that this two-staged herb therapy is a safe and effective measure for endometriosis patients with infertility (Ding and Lian, 2015).

Bushen Huoxue recipe (BSHXR, Table III) is a classic Chinese herbal prescription for nourishing the kidney and activating blood circulation, traditionally used to treat failed pregnancy and its complications (Ding *et al.*, 2018). In a meta-analysis including a total of 936 patients, it was observed that BSHXR does not significantly differ from Western medicine in terms of improving endometriosis-associated pain. However, BSHXR significantly enhanced the clinical pregnancy rate. In addition, although six trials described serious events, less adverse effects of BSHXR were reported when compared to Western medicine (Shan *et al.*, 2017).

Danefukang (DEFK, Table III) is a manufactured decoction based on CHM formula, and is currently an over-the-counter drug with indications for dysmenorrheal, menstrual disturbances (Guo *et al.*, 2010). DEFK soft extract is the first CHM formula that received a national (Chinese) drug approval number for the treatment of endometriosis (Li *et al.*, 2017). A meta-analysis of 39 randomised controlled trials enrolling 5442 patients was carried out to evaluate the efficacy and safety of DEFK soft extract in the treatment of endometriosis (Li *et al.*,

2017). The results revealed that DEFK soft extract was more efficient than gestrinone in the treatment of endometriosis and its efficacy was comparable to that of danazol and mifepristone. In terms of the relapse rate and relieving dysmenorrhea, DEFK soft extract was also as effective as gestrinone and mifepristone, and the incidence of adverse reactions was lower than that of conventional Western medicines. In conclusion, DEFK soft extract offers certain advantages in endometriosis treatment. However, because the methodological quality of the studies included in this analysis was low, rigorously designed and strictly implemented randomised controlled trials are required to further validate the efficacy of DEFK (Li *et al.*, 2017). For instance, a recent report of Zhong *et al.* (2019) analysed the efficacy of DEFK soft extract for the treatment of symptoms associated with endometriosis. A total of 174 patients with endometriosis were randomly divided into a group treated with DEFK or a control group treated with mifepristone. Both groups were treated for 3 months. The effectiveness rate was significantly superior in the DEFK group than in the mifepristone control group and treatment with DEFK resulted in a greater improvement in quality of life (improved symptoms, attenuated depression and anxiety). Moreover, DEFK treatment resulted in a decrease in the levels of the pro-inflammatory cytokines TNF- α and IL-6 and reduced CA-125 levels compared with mifepristone. In conclusion, it was suggested that DEFK is a meaningful option for patients with endometriosis (Zhong *et al.*, 2019).

Currently, the ClinicalTrials.gov website lists endometriosis as one of the diseases included in a large recruiting randomised interventional clinical trial (720 participants) comparing the CHM Xuefu-Zhuyu capsule (XFZY, Table III) administered at 2.4 g (6 capsules) three times daily for 12 weeks with placebo (Wang, 2020b). The primary outcome are changes in the Chinese Medicine patient-reported-outcome scale in patients with Qi Stagnation and Blood Stasis Syndrome, characterised by symptoms such as different types of pain, irritability or depression, insomnia and chest tightness, among others (Hu *et al.*, 2015). Pain, self-rating anxiety scale, self-rating depression scale and Quality of Life Scale are listed as secondary outcomes along with an ambitious monitoring of gene expression and biochemical parameters. Notably, XFZY has been used in CHM as a treatment of primary dysmenorrhea, indicating a possible link to endometriosis (Leem *et al.*, 2019).

Nowadays, ancient formulations of CHM and Kampo (Japanese) medicines coexist and complement each other. The herbs used in Kampo are commonly used for making decoctions and herbal preparations from a single crude ingredient or a mixture of multiple crude ingredients (Teng *et al.*, 2016). Among them, tokishakuyakusan (TSS, Table III) is a representative Kampo mixture, which is widely prescribed in Japan for patients with gynecological disorders (Nagira *et al.*, 2019). TSS has been shown to reduce the severity of primary and secondary dysmenorrhea (Oya *et al.*, 2008; Yoshino *et al.*, 2016) and to alleviate menstruation-related headaches that are refractory to conventional treatments (Akaishi *et al.*, 2019). In Balb/c mice with induced endometriosis, TSS treatment significantly reduced lesion number and size, and improved sensitivity to nociceptive stimuli. It was assumed that paeoniflorin contained in paeoniae radix is the main component responsible for the antinociceptive of TSS (Nagira *et al.*, 2019). In a recent *in vitro* study, Takeuchi *et al.* (2020) further observed that TSS has anti-inflammatory and anti-angiogenic effects on endometriosis by controlling IL-8 and VEGF secretion in endometriotic stromal cells.

Table IV CHM formulae evaluated in experimental models of endometriosis.

| Formula | Model/dosage | Action/mechanism | Reference |
|--|--|--|------------------------|
| Ping-Chong-Jiang-Ni (PCJNF) | Human ectopic endometrial stromal cells | ↓ Cell proliferation ↓ Invasion ↓ Cell migration ↑ Apoptosis | Liang et al. (2017) |
| Liu Fang (XLF) | Human ectopic and eutopic endometrial stromal cells from patients with endometriosis | ↓ Attachment ↓ Angiogenesis ↓ ICAM-1 ↓ COX-2 ↓ MMP-9 ↓ VEGF | Zhou et al. (2019) |
| Wenshen Xiaozheng Tang (WXT) | Human ectopic endometrial stromal cells | ↓ Cell proliferation ↑ Apoptosis ↓ NF-κB ↓ Cell migration | Zhang et al. (2016) |
| | Sprague-Dawley rats | ↓ Endometriotic lesion size ↓ IL-1β ↓ TNF-α ↓ ICAM-1 ↓ COX-2 ↓ MMP-9 ↑ TIMP-1 | Zhang et al. (2013b) |
| Gui-Zhi-Fu-Ling Capsule (GZFLC) | Sprague-Dawley rats | ↓ Endometriotic lesion size ↓ ICAM-1 ↓ MCP-1 ↓ CD4 ⁺ lymphocytes ↓ Natural killer cell activity | Ji et al. (2011) |
| | Sprague-Dawley rats | ↑ Apoptosis ↑ Bax ↓ Bcl-2 ↓ Cell proliferation | Hu et al. (2014) |
| | Wistar rats | ↓ GLUT-4 ↓ VEGF ↓ TGF-β | Zhou et al. (2018) |
| Sanjie Zhentong capsule (SJZTC) | Lewis rats | ↓ Endometriotic lesion size ↓ VEGF ↓ TNF-α | Zou et al. (2013) |
| Xiaochaihu Tang (XCHT) | Sprague-Dawley rats | ↓ MMP-2 ↓ MMP-9 | Jiao et al. (2013) |
| Hua Yu Xiao Zheng (HYXZ) | Sprague-Dawley rats | ↓ Endometriotic lesion size ↓ Angiogenesis ↓ VEGF ↓ Angiopoietin-2 | Chen and Gong (2017) |
| Shaofu Zhuyu Decoction (SZD) | Sprague-Dawley rats | ↓ Endometriotic lesion size ↓ Cell proliferation ↑ Apoptosis ↓ Angiogenesis ↓ HIF-1α | Zhu et al. (2018) |
| Jiawei-Foshou-San (JFS) | Sprague-Dawley rats | ↓ Endometriotic lesion size ↓ Invasion and metastasis Regulation of epithelial-mesenchymal transformation | Chen et al. (2018b) |
| Tokishakuyakusan (TSS) | Balb/c mice | ↓ Endometriotic lesion number ↓ Endometriotic lesion size ↓ Endometriosis-pain | Nagira et al. (2019) |
| | Human ectopic endometrial stromal cells | ↓ IL-8 ↓ VEGF | Takeuchi et al. (2020) |

Bax, Bcl-2-associated X protein; Bcl-2, B cell lymphoma-2; Glut-4, glucose transporter type 4; HIF-1α, hypoxia-inducible factor-1; TGF-β, transforming growth factor β.

Summary

As summarised in Table IV, different CHM formulations have been shown to target multiple cellular processes that are crucially involved in the pathogenesis of endometriosis, such as inflammation, oxidative stress and apoptosis. Hence, they have also been demonstrated to effectively inhibit the development and growth of endometriotic lesions in different animal models of the disease. Additional clinical findings indicate that CHM may be particularly beneficial to treat endometriosis-associated symptoms, such as pain. In this context, all published studies reported that CHM has at least the same efficacy as the standard Western medicine with fewer side effects. However, the underlying mechanisms remain elusive so far. Moreover, it should be noted that in the past most clinical trials on the use of traditional Chinese medicine to treat endometriosis have been judged to be of poor quality due to substantial flaws in evaluating outcome measures, follow-up, diagnosis, data analysis, reporting of adverse effects and randomisation (Guo et al., 2010). Indeed, in over 850 CHM studies of endometriosis analysed by Guo et al. (2010), only 18.0% achieved evidence Level II, and none had a Jadad score >3 , which is considered the minimum standard for inclusion in a systematic review or meta-analysis. Hence, additional convincing evidence is needed to prove the suitability of CHM for endometriosis treatment. If this succeeds, the integration of CHM into Western medicine could be a promising approach in the future multi-modal management of endometriosis.

Conclusions

The herein described preclinical and clinical studies indicate that natural plant-derived agents represent promising candidates for the development of novel treatment strategies in the management of endometriosis. They typically exhibit a pleiotropic action profile, which simultaneously targets fundamental processes in the complex pathophysiology of the disease, such as proliferation, apoptosis, inflammation, ROS formation and angiogenesis. Hence, they may help to prevent escape mechanisms, as they are frequently observed in response to targeted therapy. Moreover, they may induce less severe side effects when compared to conventional anti-oestrogenic compounds. In addition, they bear the chance to introduce affordable therapeutics into the market, which is a big issue in all countries given the continuously rising worldwide healthcare costs.

However, there are still many hurdles to overcome for the successful implementation of natural treatment strategies in daily clinical practice. The heterogenous and often combinatorial nature of the natural compounds makes it difficult to differentiate cause from effect in the readouts of numerous experimental studies, and the exact mechanism of action is often unknown. Moreover, pharmacokinetics and pharmacodynamics have not been carefully addressed in many of the preclinical studies, and herb–drug interactions need to be carefully studied when the natural compounds are combined with conventional pharmacotherapy. Indeed, their efficiency and safety has to be proven in comparison to conventional therapeutic approaches in randomised controlled multi-centre trials. However, for this purpose it is necessary to establish standardised protocols for the GMP-compliant generation of well-defined plant extracts. In this context, it should also be considered that many herbs or herbal compounds may not be patentable or

their patents may have expired a long time ago. Hence, there is no incentive for pharmaceutical industry to drive the development in this field or even to conduct expensive clinical trials. Another important issue is the fact that most phytochemicals exhibit a poor stability and bioavailability in their natural form and, thus, may primarily serve as lead compounds for the development of novel drugs with improved pharmacokinetic profiles (Phan et al., 2018). Finally, it should be noted that natural-based therapies, particularly CHM, primarily focus on the improvement of endometriosis-related symptoms but not on the cure of the disease. This may also be a major reason why even within China itself CHM is not accepted and approved by all gynaecologists and patients (Guo et al., 2010). Moreover, it is unlikely that natural-based therapies will ever serve as a stand-alone therapy. However, they bear great potential as an integral part of future multi-modal treatment concepts for endometriosis. This is underscored by the rapidly growing interest and demand for these therapies by not only endometriosis patients but also their attending physicians.

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Authors' roles

G.F.M. formulated the primary draft of the manuscript and primarily carried out the PubMed searches, analysed the data and drafted the tables, M.G. supervised the conception, design and development of all aspects of the article, carried out the Clinicaltrials.gov searches, designed the figures and was involved in the data analysis and interpretation. M.W.L. made substantial contributions to design, analysis and interpretation of data. Development of the article, review, editing and final approval of the final article was by G.F.M., M.G. and M.W.L.

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Conflict of interest

All authors declare that no conflict of interest exists.

References

- Abdel-Sattar EA, Mouneir SM, Asaad GF, Abdallah HM. Protective effect of *Calligonum comosum* on haloperidol-induced oxidative stress in rat. *Toxicol Ind Health* 2014;30:147–153.
- Abdo W, Hirata A, Shukry M, Kamal T, Abdel-Sattar E, Mahrous E, Yanai T. *Calligonum comosum* extract inhibits diethylnitrosamine-induced hepatocarcinogenesis in rats. *Oncol Lett* 2015;10:716–722.

Akaishi T, Takayama S, Ohsawa M, Kikuchi A, Arita R, Nakashima I, Aoki M, Ishii T. Successful treatment of intractable menstrual migraine with the traditional herbal medicine tokishakuyakusan. *J Gen Fam Med* 2019; **20**:118–121.

Amaya SC, Savaris RF, Filipovic CJ, Wise JD, Hestermann E, Young SL, Lessey BA. Resveratrol and endometrium: a closer look at an active ingredient of red wine using *in vivo* and *in vitro* models. *Reprod Sci* 2014; **21**:1362–1369.

Arablou T, Delbandi AA, Khodaverdi S, Arefi S, Kolahdouz-Mohammadi R, Heidari S, Mohammadi T, Aryaeian N. Resveratrol reduces the expression of insulin-like growth factor-1 and hepatocyte growth factor in stromal cells of women with endometriosis compared with nonendometriotic women. *Phyther Res* 2019; **33**:1044–1054.

Arablou T, Kolahdouz-Mohammadi R. Curcumin and endometriosis: review on potential roles and molecular mechanisms. *Biomed Pharmacother* 2018; **97**:91–97.

ASRM. Revised American Society for Reproductive Medicine classification of endometriosis: 1996. *Fertil Steril* 1997; **67**:817–821.

Attar E, Bulun SE. Aromatase and other steroidogenic genes in endometriosis: translational aspects. *Hum Reprod Update* 2006; **12**:49–56.

Attar E, Tokunaga H, Imlir G, Yilmaz MB, Redwine D, Putman M, Gurates B, Attar R, Yaegashi N, Hales DB et al. Prostaglandin E2 via steroidogenic factor-1 coordinately regulates transcription of steroidogenic genes necessary for estrogen synthesis in endometriosis. *J Clin Endocrinol Metab* 2009; **94**:623–631.

Aznaurova YB, Zhumataev MB, Roberts TK, Aliper AM, Zhavoronkov AA. Molecular aspects of development and regulation of endometriosis. *Reprod Biol Endocrinol* 2014; **12**:50.

Badria FA, Ameen M, Akl MR. Evaluation of cytotoxic compounds from *Calligonum comosum* L. growing in Egypt. *Z Naturforsch C* 2007; **62**:656–660.

Bayoglu Tekin Y, Guven S, Kirbas A, Kalkan Y, Tumkaya L, Guvendag Guven ES. Is resveratrol a potential substitute for leuprolide acetate in experimental endometriosis? *Eur J Obstet Gynecol Reprod Biol* 2015; **184**:1–6.

Becker CM, D'Amato RJ. Angiogenesis and antiangiogenic therapy in endometriosis. *Microvasc Res* 2007; **74**:121–130.

Bina F, Soleymani S, Toliat T, Hajimahmoodi M, Tabarrai M, Abdollahi M, Rahimi R. Plant-derived medicines for treatment of endometriosis: a comprehensive review of molecular mechanisms. *Pharmacol Res* 2019; **139**:76–90.

Bisht K, Wagner KH, Bulmer AC. Curcumin, resveratrol and flavonoids as anti-inflammatory, cyto- and DNA-protective dietary compounds. *Toxicology* 2010; **278**:88–100.

Bouaziz J, Bar On A, Seidman DS, Soriano D. The clinical significance of endocannabinoids in endometriosis pain management. *Cannabis Cannabinoid Res* 2017; **2**:72–80.

Bourlev V, Volkov N, Pavlovitch S, Lets N, Larsson A, Olovsson M. The relationship between microvessel density, proliferative activity and expression of vascular endothelial growth factor-A and its receptors in eutopic endometrium and endometriotic lesions. *Reproduction* 2006; **132**:501–509.

Boyaji S, Merkow J, Elman RNM, Kaye AD, Yong RJ, Urman RD. The role of cannabidiol (CBD) in chronic pain management: an assessment of current evidence. *Curr Pain Headache Rep* 2020; **24**:4.

Bruner-Tran KL, Osteen KG, Taylor HS, Sokalska A, Haines K, Duleba AJ. Resveratrol inhibits development of experimental endometriosis *in vivo* and reduces endometrial stromal cell invasiveness *in vitro*. *Biol Reprod* 2011; **84**:106–112.

Bulun SE, Monsivais D, Kakinuma T, Furukawa Y, Bernardi L, Pavone ME, Dyson M. Molecular biology of endometriosis: from aromatase to genomic abnormalities. *Semin Reprod Med* 2015; **33**:220–224.

Burks-Wicks C, Cohen M, Fallbacher J, Taylor RN, Wieser F. A Western primer of Chinese herbal therapy in endometriosis and infertility. *Women's Heal* 2005; **1**:447–463.

Burney RO, Giudice LC. Pathogenesis and pathophysiology of endometriosis. *Fertil Steril* 2012; **98**:511–519.

Cao H, Wei YX, Zhou Q, Zhang Y, Guo XP, Zhang J. Inhibitory effect of curcumin in human endometriosis endometrial cells via downregulation of vascular endothelial growth factor. *Mol Med Rep* 2017a; **16**:5611–5617.

Cao Y, Ye Q, Zhuang M, Xie S, Zhong R, Cui J, Zhou J, Zhu Y, Zhang T, Cao L. Ginsenoside Rg3 inhibits angiogenesis in a rat model of endometriosis through the VEGFR-2-mediated PI3K/Akt/mTOR signaling pathway. *PLoS One* 2017b; **12**:e0186520.

Cao Y, Zhuang MF, Yang Y, Xie SW, Cui JG, Cao L, Zhang TT, Zhu Y. Preliminary study of quercetin affecting the hypothalamic-pituitary-gonadal axis on rat endometriosis model. *Evid Based Complement Altern Med* 2014; **2014**:781684.

Capobianco A, Rovere-Querini P. Endometriosis, a disease of the macrophage. *Front Immunol* 2013; **4**:9.

Cenksoy PO, Oktem M, Erdem O, Karakaya C, Cenksoy C, Erdem A, Guner H, Karabacak O. A potential novel treatment strategy: inhibition of angiogenesis and inflammation by resveratrol for regression of endometriosis in an experimental rat model. *Gynecol Endocrinol* 2015; **31**:219–224.

Chen D, Wan SB, Yang H, Yuan J, Chan TH, Dou QP. EGCG, green tea polyphenols and their synthetic analogs and prodrugs for human cancer prevention and treatment. *Adv Clin Chem* 2011a; **53**:155–177.

Chen X, Yu J, Shi J. Management of diabetes mellitus with puerarin, a natural isoflavone from *Pueraria lobata*. *Am J Chin Med* 2018a; **46**:1771–1789.

Chen Y, Chen C, Shi S, Han J, Wang J, Hu J, Liu Y, Cai Z, Yu C. Endometriotic implants regress in rat models treated with puerarin by decreasing estradiol level. *Reprod Sci* 2011b; **18**:886–891.

Chen Y, Wei J, Zhang Y, Sun W, Li Z, Wang Q, Xu X, Li C, Li P. Anti-endometriosis mechanism of Jiawei Foshou San based on network pharmacology. *Front Pharmacol* 2018b; **9**:811.

Chen ZZ, Gong X. Effect of Hua Yu Xiao Zheng decoction on the expression levels of vascular endothelial growth factor and angiopoietin-2 in rats with endometriosis. *Exp Ther Med* 2017; **14**:5743–5750.

Cheng W, Chen L, Yang S, Han J, Zhai D, Ni J, Yu C, Cai Z. Puerarin suppresses proliferation of endometriotic stromal cells partly via the MAPK signaling pathway induced by 17 β -estradiol-BSA. *PLoS One* 2012; **7**:e45529.

Cho S, Choi YS, Jeon YE, Im KJ, Choi YM, Yim SY, Kim H, Seo SK, Lee BS. Expression of vascular endothelial growth factor (VEGF) and its soluble receptor-1 in endometriosis. *Microvasc Res* 2012; **83**:237–242.

Chowdhury I, Banerjee S, Driss A, Xu W, Mehrabi S, Nezhat C, Sidell N, Taylor RN, Thompson WE. Curcumin attenuates proangiogenic and proinflammatory factors in human eutopic endometrial stromal cells through the NF-κB signaling pathway. *J Cell Physiol* 2019; **234**:6298–6312.

Davatgaran-Taghipour Y, Masoomzadeh S, Farzaei MH et al. Polyphenol nanoformulations for cancer therapy: experimental evidence and clinical perspective. *Int J Nanomed* 2017; **12**:2689–2702.

Della Valle V. *Uncaria tomentosa*. *G Ital Dermatol Venereol* 2017; **152**: 651–657.

Di Costanzo A, Angelico R. Formulation strategies for enhancing the bioavailability of silymarin: the state of the art. *Molecules* 2019; **24**: 2155.

Ding J, Tan X, Song K, Ma W, Xiao J, Song Y, Zhang M. Bushen Huoxue recipe alleviates implantation loss in mice by enhancing estrogen–progesterone signals and promoting decidual angiogenesis through FGF2 during early pregnancy. *Front Pharmacol* 2018; **9**:437.

Ding Z, Lian F. Traditional Chinese medical herbs staged therapy in infertile women with endometriosis: a clinical study. *Int J Clin Exp Med* 2015; **8**:14085–14089.

Dmowski WP, Ding J, Shen J, Rana N, Fernandez BB, Braun DP. Apoptosis in endometrial glandular and stromal cells in women with and without endometriosis. *Hum Reprod* 2001; **16**:1802–1808.

Dong-Soon IM, Seung-Yeol NAH. Yin and Yang of ginseng pharmacology: ginsenosides vs gintonin. *Acta Pharmacol Sin* 2013; **34**: 1367–1373.

Dull AM, Moga MA, Dimienescu OG, Sechel G, Burtea V, Anastasiu CV. Therapeutic approaches of resveratrol on endometriosis via anti-inflammatory and anti-angiogenic pathways. *Molecules* 2019; **24**:667.

Ergenoğlu AM, Yeniel AÖ, Erbaş O, Aktuğ H, Yıldırım N, Ulukış M, Taskiran D. Regression of endometrial implants by resveratrol in an experimentally induced endometriosis model in rats. *Reprod Sci* 2013; **20**:1230–1236.

Erowele GI, Kalejaiye AO. Pharmacology and therapeutic uses of cat's claw. *Am J Heal Pharm* 2009; **66**:992–995.

Escudero-Lara A, Argerich J, Cabañero D, Maldonado R. Disease-modifying effects of natural Δ9-tetrahydrocannabinol in endometriosis-associated pain. *eLife* 2020; **9**:e50356.

Fang RC, Tsai YT, Lai JN, Yeh CH, Wu CT. The traditional Chinese medicine prescription pattern of endometriosis patients in Taiwan: a population-based study. *Evid Based Complement Altern Med* 2012; **2012**:591391.

Ferella L, Bastón JI, Bilotas MA, Singla JJ, González AM, Olivares CN, Meresman GF. Active compounds present in *Rosmarinus officinalis* leaves and *Scutellaria baicalensis* root evaluated as new therapeutic agents for endometriosis. *Reprod Biomed Online* 2018; **37**:769–782.

Flower A, Liu JP, Chen S, Lewith G, Little P. Chinese herbal medicine for endometriosis. *Cochrane Database Syst Rev* 2012; **2012**: CD006568.

Freund RRA, Gobrecht P, Fischer D, Arndt HD. Advances in chemistry and bioactivity of parthenolide. *Nat Prod Rep* 2020; **37**: 541–565.

Garcia Cinca, D. *Effect of Cannabinoid (THC/CBD 50%) on Hyperalgesia in Patients with Deep Endometriosis (EdomTHC)*. Bethesda, MD, USA: U.S. National Library of Medicine, 2020. <https://clinicaltrials.gov/ct2/show/NCT03875261> (21 August 2020, date last accessed).

Gazvani R, Templeton A. Peritoneal environment, cytokines and angiogenesis in the pathophysiology of endometriosis. *Reproduction* 2002; **123**:217–226.

Ghantous A, Sinjab A, Herceg Z, Darwiche N. Parthenolide: from plant shoots to cancer roots. *Drug Discov Today* 2013; **18**:894–905.

Greene AD, Lang SA, Kendzierski JA, Sroga-Rios JM, Herzog TJ, Burns KA. Endometriosis: where are we and where are we going? *Reproduction* 2016; **152**:63–78.

Groothuis PG, Nap AW, Winterhager E, Grümmer R. Vascular development in endometriosis. *Angiogenesis* 2005; **8**:147–156.

Guo SW. Recurrence of endometriosis and its control. *Hum Reprod Update* 2009; **15**:441–461.

Guo SW, He W, Zhao T, Liu X, Zhang T. Clinical trials and trial-like studies on the use of traditional Chinese medicine to treat endometriosis. *Expert Rev Obstet Gynecol* 2010; **5**:533–555.

Guo SW, Martin DC. The perioperative period: a critical yet neglected time window for reducing the recurrence risk of endometriosis? *Hum Reprod* 2019; **34**:1858–1865.

Hatamipour M, Ramezani M, Tabassi SAS, Johnston TP, Sahebkar A. Demethoxycurcumin: a naturally occurring curcumin analogue for treating non-cancerous diseases. *J Cell Physiol* 2019; **234**: 19320–19330.

Hey-Cunningham AJ, Peters KM, Zevallos HB-V, Berbic M, Markham R, Fraser IS. Angiogenesis, lymphangiogenesis and neurogenesis in endometriosis. *Front Biosci* 2013; **5**:1033–1056.

Hosseinabadi T, Lorigooini Z, Tabarzad M, Salehi B, Rodrigues CF, Martins N, Sharifi-Rad J. Silymarin antiproliferative and apoptotic effects: insights into its clinical impact in various types of cancer. *Phyther Res* 2019; **33**:2849–2861.

Hu B, Wang SS, Du Q. Traditional Chinese medicine for prevention and treatment of hepatocarcinoma: from bench to bedside. *World J Hepatol* 2015; **7**:1209–1232.

Hu C, Wang Z, Pang Z, Lu W, Cai X, Yang J, Wang D, Cao P. Guizhi Fuling capsule, an ancient Chinese formula, attenuates endometriosis in rats via induction of apoptosis. *Climacteric* 2014; **17**: 410–416.

Huang R, Chen S, Zhao M, Li Z, Zhu L. Ginsenoside Rg3 attenuates endometriosis by inhibiting the viability of human ectopic endometrial stromal cells through the nuclear factor-κappaB signaling pathway. *J Gynecol Obstet Hum Reprod* 2020; **49**:101642.

Intagliata S, Modica MN, Santagati LM, Montenegro L. Strategies to improve resveratrol systemic and topical bioavailability: an update. *Antioxidants (Basel)* 2019; **8**:244.

Jamwal R. Bioavailable curcumin formulations: a review of pharmacokinetic studies in healthy volunteers. *J Integr Med* 2018; **16**: 367–374.

Jana S, Chakravarty B, Chaudhury K. Letrozole and curcumin loaded-PLGA nanoparticles: a therapeutic strategy for endometriosis. *J Nanomed Biother Discov* 2014; **4**:123.

Jana S, Paul S, Swarnakar S. Curcumin as anti-endometriotic agent: implication of MMP-3 and intrinsic apoptotic pathway. *Biochem Pharmacol* 2012; **83**:797–804.

Jelodar G, Azimifar A. Evaluation of serum cancer antigen 125, resistin, leptin, homocysteine, and total antioxidant capacity in rat

model of endometriosis treated with curcumin. *Physiol Rep* 2019; **7**: e14016.

Ji M, Liu Y, Yang S, Zhai D, Zhang D, Bai L, Wang Z, Yu J, Yu C, Cai Z. Puerarin suppresses proliferation of endometriotic stromal cells in part via differential recruitment of nuclear receptor coregulators to estrogen receptor- α . *J Steroid Biochem Mol Biol* 2013; **138**:421–426.

Ji X, Gao J, Cai X, Lu W, Hu C, Wang Z, Cheng X, Gu Z, Wan G, Zhang S et al. Immunological regulation of Chinese herb Guizhi Fuling Capsule on rat endometriosis model. *J Ethnopharmacol* 2011; **134**:624–629.

Jiang CH, Sun TL, Xiang DX, Wei SS, Li WQ. Anticancer activity and mechanism of xanthohumol: a prenylated flavonoid from hops (*Humulus lupulus L.*). *Front Pharmacol* 2018; **9**:530.

Jiao L, Qi X, Lu G, Zhang Q, Zhang C, Gao J. Effect of traditional Chinese medicine (Xiaochaihu Tang) on the expression of MMP-2 and MMP-9 in rats with endometriosis. *Exp Ther Med* 2013; **6**: 1385–1389.

Jin Z, Huang J, Zhu Z. Baicalein reduces endometriosis by suppressing the viability of human endometrial stromal cells through the nuclear factor- κ B pathway *in vitro*. *Exp Ther Med* 2017; **14**: 2992–2998.

Jouhari S, Mohammadzadeh A, Soltanghorae H, Mohammadi Z, Khazali S, Mirzadegan E, Lakpour N, Fatemi F, Zafardoust S, Mohazzab A et al. Effects of silymarin, cabergoline and letrozole on rat model of endometriosis. *Taiwan J Obstet Gynecol* 2018; **57**: 830–835.

Ju J, Lu G, Lambert JD, Yang CS. Inhibition of carcinogenesis by tea constituents. *Semin Cancer Biol* 2007; **17**:395–402.

Kao LC, Germeyer A, Tulac S, Lobo S, Yang JP, Taylor RN, Osteen K, Lessey BA, Giudice LC. Expression profiling of endometrium from women with endometriosis reveals candidate genes for disease-based implantation failure and infertility. *Endocrinology* 2003; **144**:2870–2881.

Kiani K, Rudzitis-Auth J, Scheuer C, Movahedin M, Sadati Lamardi SN, Malekafzali Ardakani H, Becker V, Moini A, Aflatoonian R, Ostad SN et al. *Calligonum comosum* (Escanbil) extract exerts anti-angiogenic, anti-proliferative and anti-inflammatory effects on endometriotic lesions. *J Ethnopharmacol* 2019; **239**:111918.

Kim JH, Woo JH, Kim HM, Oh MS, Jang DS, Choi JH. Anti-endometriotic effects of pueraria flower extract in human endometriotic cells and mice. *Nutrients* 2017; **9**:212.

Kim KH, Lee EN, Park JK, Lee JR, Kim JH, Choi HJ, Kim BS, Lee HW, Lee KS, Yoon S. Curcumin attenuates TNF- α -induced expression of intercellular adhesion molecule-1, vascular cell adhesion molecule-1 and proinflammatory cytokines in human endometriotic stromal cells. *Phyther Res* 2012; **26**:1037–1047.

Kim KH, Park JK, Choi YW, Kim YH, Lee EN, Lee JR, Kim HS, Baek SY, Kim BS, Lee KS et al. Hexane extract of aged black garlic reduces cell proliferation and attenuates the expression of ICAM-1 and VCAM-1 in TNF- α -activated human endometrial stromal cells. *Int J Mol Med* 2013; **32**:67–78.

Kizilay G, Uz YH, Seren G, Ulucam CE, Yilmaz A, Cukur Z, Kayisli UA. *In vivo* effects of curcumin and deferoxamine in experimental endometriosis. *Adv Clin Exp Med* 2017; **26**:207–213.

Kundu JK, Surh YJ. Cancer chemopreventive and therapeutic potential of resveratrol: mechanistic perspectives. *Cancer Lett* 2008; **269**: 243–261.

Laganà AS, Garzon S, Götte M, Viganò P, Franchi M, Ghezzi F, Martin DC. The pathogenesis of endometriosis: molecular and cell biology insights. *Int J Mol Sci* 2019; **20**:5615.

Laschke MW, Menger MD. Basic mechanisms of vascularization in endometriosis and their clinical implications. *Hum Reprod Update* 2018; **24**:207–224.

Laschke MW, Menger MD. *In vitro* and *in vivo* approaches to study angiogenesis in the pathophysiology and therapy of endometriosis. *Hum Reprod Update* 2007; **13**:331–342.

Laschke MW, Schwender C, Scheuer C, Vollmar B, Menger MD. Epigallocatechin-3-gallate inhibits estrogen-induced activation of endometrial cells *in vitro* and causes regression of endometriotic lesions *in vivo*. *Hum Reprod* 2008; **23**:2308–2318.

Laschke MW, Van Oijen AEV, Scheuer C, Menger MD. *In vitro* and *in vivo* evaluation of the anti-angiogenic actions of 4-hydroxybenzyl alcohol. *Br J Pharmacol* 2011; **163**:835–844.

Laschke MW, Vorsterman Van Oijen AE, Körbel C, Scheuer C, Menger MD. 4-Hydroxybenzyl alcohol: a novel inhibitor of tumor angiogenesis and growth. *Life Sci* 2013; **93**:44–50.

Lee W, Ku SK, Bae JS. Anti-inflammatory effects of baicalin, baicalein, and wogonin *in vitro* and *in vivo*. *Inflammation* 2014; **38**: 110–125.

Leem J, Jo J, Kwon CY, Lee H, Park KS, Lee JM. Herbal medicine (Hyebulbuchukeo-tang or Xuefu Zhuyu decoction) for treating primary dysmenorrhea: a systematic review and meta-analysis of randomized controlled trials. *Medicine (Baltimore)* 2019; **98**: e14170.

Lessey BA, Lebovic DI, Taylor RN. Eutopic endometrium in women with endometriosis: ground zero for the study of implantation defects. *Semin Reprod Med* 2013; **31**:109–124.

Li Y, Chen H, Yang S, Xie G. Decreased expression of aromatase in the Ishikawa and RL95-2 cells by the isoflavone, puerarin, is associated with inhibition of c-Jun expression and AP-1 activity. *Food Chem Toxicol* 2008; **46**:3671–3676.

Li Y, Li T, Song S. Evaluation of efficacy and safety of Dan'e-fukang soft extract in the treatment of endometriosis: a meta-analysis of 39 randomized controlled trials enrolling 5442 patients. *Evid Based Complement Altern Med* 2017; **2017**:9767391.

Liang RN, Li PS, Zou Y, Liu YL, Jiang Z, Liu Z, Fan P, Xu L, Peng JH, Sun XY. Ping-Chong-Jiang-Ni formula induces apoptosis and inhibits proliferation of human ectopic endometrial stromal cells in endometriosis via the activation of JNK signaling pathway. *Evid Based Complement Altern Med* 2017; **2017**:6489427.

Liu H, Lang JH. Is abnormal eutopic endometrium the cause of endometriosis? The role of eutopic endometrium in pathogenesis of endometriosis. *Med Sci Monit* 2011; **17**:RA92–9.

Liu J, Zhang Q, Li R, Wei S, Huang C, Gao Y, Pu X. The traditional uses, phytochemistry, pharmacology and toxicology of *Cinnamomi ramulus*: a review. *J Pharm Pharmacol* 2020; **72**:319–342.

Liu JP, Yang H, Xia Y, Cardini F. Herbal preparations for uterine fibroids. *Cochrane Database Syst Rev* 2013; **2013**:CD005292.

Li-Weber M. New therapeutic aspects of flavones: the anticancer properties of *Scutellaria* and its main active constituents wogonin, baicalein and baicalin. *Cancer Treat Rev* 2009; **35**:57–68.

López-Jiménez A, García-Caballero M, Medina MÁ, Quesada AR. Anti-angiogenic properties of carnosol and carnosic acid, two major dietary compounds from rosemary. *Eur J Nutr* 2013; **52**:85–95.

Lousse JC, Defrre S, Colette S, Van Langendonckt A, Donnez J. Expression of eicosanoid biosynthetic and catabolic enzymes in peritoneal endometriosis. *Hum Reprod* 2010; **25**:734–741.

Lousse JC, Van Langendonckt A, Defrre S, Ramos RG, Colette S, Donnez J. Peritoneal endometriosis is an inflammatory disease. *Front Biosci* 2012; **4**:23–40.

Ma Z, Wang N, He H, Tang X. Pharmaceutical strategies of improving oral systemic bioavailability of curcumin for clinical application. *J Control Release* 2019; **316**:359–380.

Machado DE, Rodrigues-Baptista KC, Alessandra-Perini J, De Moura RS, Dos Santos TA, Pereira KG, Da Silva YM, Souza PJ, Nasciutti LE, Perini JA. *Euterpe oleracea* extract (Açaí) is a promising novel pharmacological therapeutic treatment for experimental endometriosis. *PLoS One* 2016; **11**:e0166059.

Maia H, Haddad C, Pinheiro N, Casoy J. Advantages of the association of resveratrol with oral contraceptives for management of endometriosis-related pain. *Int J Womens Health* 2012; **4**:543–549.

Margalioth EJ, Ben-Chetrit A, Gal M, Eldar-Geva T. Investigation and treatment of repeated implantation failure following IVF-ET. *Hum Reprod* 2006; **21**:3036–3043.

Matsuzaki S, Darcha C. Antifibrotic properties of epigallocatechin-3-gallate in endometriosis. *Hum Reprod* 2014; **29**:1677–1687.

Mendes da Silva D, Gross LA, Neto E de PG, Lessey BA, Savaris RF. The use of resveratrol as an adjuvant treatment of pain in endometriosis: a randomized clinical trial. *J Endocr Soc* 2017; **1**:359–369.

Meresman GF, Vighi S, Buquet RA, Contreras-Ortiz O, Tesone M, Rumi LS. Apoptosis and expression of Bcl-2 and Bax in eutopic endometrium from women with endometriosis. *Fertil Steril* 2000; **74**:760–766.

Moore J, Yousef M, Tsiani E. Anticancer effects of rosemary (*Rosmarinus officinalis* L.) extract and rosemary extract polyphenols. *Nutrients* 2016; **8**:731.

Nagira K, Taniguchi F, Nakamura K, Tokita Y, Tsuchiya N, Khine YM, Harada T. Tokishakuyakusan, a Kampo medicine, attenuates endometriosis-like lesions and hyperalgesia in murine with endometriosis-like symptoms. *Am J Reprod Immunol* 2019; **82**:e13182.

Nahari E, Razi M. Silymarin amplifies apoptosis in ectopic endometrial tissue in rats with endometriosis; implication on growth factor GDNF, ERK1/2 and Bcl-6b expression. *Acta Histochem* 2018; **120**:757–767.

Nasu K, Yuge A, Tsuno A, Nishida M, Narahara H. Involvement of resistance to apoptosis in the pathogenesis of endometriosis. *Histol Histopathol* 2009; **24**:1181–1192.

Neto JN, Coelho TM, Aguiar GC, Carvalho LR, De Araújo AGP, Girão MJBC, Schor E. Experimental endometriosis reduction in rats treated with *Uncaria tomentosa* (cat's claw) extract. *Eur J Obstet Gynecol Reprod Biol* 2011; **154**:205–208.

Oya A, Oikawa T, Nakai A, Takeshita T, Hanawa T. Clinical efficacy of Kampo medicine (Japanese traditional herbal medicine) in the treatment of primary dysmenorrhea. *J Obstet Gynaecol Res* 2008; **34**:898–908.

Park C, Park S, Chung YH, Kim GY, Choi YWYH, Kim BW, Choi YWYH. Induction of apoptosis by a hexane extract of aged black garlic in the human leukemic U937 cells. *Nutr Res Pract* 2014; **8**:132–137.

Park H, Kim S, Kim S, Song Y, Seung K, Hong D, Khang G, Lee D. Antioxidant and anti-inflammatory activities of hydroxybenzyl alcohol releasing biodegradable polyoxalate nanoparticles. *Biomacromolecules* 2010; **11**:2103–2108.

Park S, Lim W, Bazer FW, Song G. Apigenin induces ROS-dependent apoptosis and ER stress in human endometriosis cells. *J Cell Physiol* 2018; **233**:3055–3065.

Park S, Lim W, Bazer FW, Whang KY, Song G. Quercetin inhibits proliferation of endometriosis regulating cyclin D1 and its target microRNAs *in vitro* and *in vivo*. *J Nutr Biochem* 2019; **63**:87–100.

Perricos, A. Medical University of Vienna. *Flexofytol® for the Treatment of Endometriosis-Associated Pain (ENDOFLEX)*. Bethesda, MD, USA: U.S. National Library of Medicine, 2020. <https://clinicaltrials.gov/ct2/show/NCT04150406> (21 August 2020, date last accessed).

Petiwal SM, Johnson JJ. Diterpenes from rosemary (*Rosmarinus officinalis*): defining their potential for anti-cancer activity. *Cancer Lett* 2015; **367**:93–102.

Phan MAT, Paterson J, Bucknall M, Arcot J. Interactions between phytochemicals from fruits and vegetables: effects on bioactivities and bioavailability. *Crit Rev Food Sci Nutr* 2018; **58**:1310–1329.

Polka D, Podsedek A, Koziołkiewicz M. Comparison of chemical composition and antioxidant capacity of fruit, flower and bark of *Viburnum opulus*. *Plant Foods Hum Nutr* 2019; **74**:436–442.

Qin X, Liu Y, Feng Y, Jiang J. Ginsenoside Rf alleviates dysmenorrhea and inflammation through the BDNF-TrkB-CREB pathway in a rat model of endometriosis. *Food Funct* 2019; **10**:244–249.

Rauf A, Imran M, Butt MS, Nadeem M, Peters DG, Mubarak MS. Resveratrol as an anti-cancer agent: a review. *Crit Rev Food Sci Nutr* 2018; **58**:1428–1447.

Reis FM, Petraglia F, Taylor RN. Endometriosis: hormone regulation and clinical consequences of chemotaxis and apoptosis. *Hum Reprod Update* 2013; **19**:406–418.

Reyes-Farias M, Carrasco-Pozo C. The anti-cancer effect of quercetin: molecular implications in cancer metabolism. *Int J Mol Sci* 2019; **20**:3177.

Ricci AG, Olivares CN, Bilotas MA, Bastón JL, Singla JJ, Meresman GF, Barañao RI. Natural therapies assessment for the treatment of endometriosis. *Hum Reprod* 2013; **28**:178–188.

Ricci AG, Olivares CN, Bilotas MA, Meresman GF, Barañao RI. Effect of vascular endothelial growth factor inhibition on endometrial implant development in a murine model of endometriosis. *Reprod Sci* 2011; **18**:614–622.

Ried K. Chinese herbal medicine for female infertility: an updated meta-analysis. *Complement Ther Med* 2015; **23**:116–128.

Ried K, Stuart K. Efficacy of traditional Chinese herbal medicine in the management of female infertility: a systematic review. *Complement Ther Med* 2011; **19**:319–331.

Rosa-e-Silva JC, Garcia SB, de Sá Rosa-e-Silva ACJ, Cândido-dos-Reis FJ, Poli-Neto OB, Ferriani RA, Nogueira AA. Increased cell proliferation in experimentally induced endometriosis in rabbits. *Fertil Steril* 2010; **93**:1637–1642.

Roshangar L, Abdollahifard S, Majdi A, Zarrintan A, Ghasemzade A, Farzadi L, Soleimani Rad S, Soleimani Rad J. Study of ultrastructure and apoptosis in the endometrium of women with or without endometriosis. *Iran J Reprod Med* 2013; **11**:399–404.

Rudzitis-Auth J, Krbel C, Scheuer C, Menger MD, Laschke MW. Xanthohumol inhibits growth and vascularization of developing endometriotic lesions. *Hum Reprod* 2012; **27**:1735–1744.

Rudzitis-Auth J, Menger MD, Laschke MW. Resveratrol is a potent inhibitor of vascularization and cell proliferation in experimental endometriosis. *Hum Reprod* 2013; **28**:1339–1347.

Salehi B, Venditti A, Sharifi-Rad M, Kregiel D, Sharifi-Rad J, Durazzo A, Lucarini M, Santini A, Souto EB, Novellino E et al. The therapeutic potential of Apigenin. *Int J Mol Sci* 2019; **20**:1305.

Saltan G, Süntar I, Ozbilgin S, İlhan M, Demirel MA, Oz BE, Keleş H, Akkol EK. *Viburnum opulus* L.: a remedy for the treatment of endometriosis demonstrated by rat model of surgically-induced endometriosis. *J Ethnopharmacol* 2016; **193**:450–455.

Sampson JA. Peritoneal endometriosis due to the menstrual dissemination of endometrial tissue into the peritoneal cavity. *Am J Obstet Gynecol* 1927; **14**:422–469.

Santulli P, Marcellin L, Menard S, Thubert T, Khoshnood B, Gayet V, Goffinet F, Ancel P-Y, Chapron C. Increased rate of spontaneous miscarriages in endometriosis-affected women. *Hum Reprod* 2016; **31**:1014–1023.

Sarıözkan S, Türk G, Eken A, Bayram LC, Baldemir A, Doğan G. Gilaburu (*Viburnum opulus* L.) fruit extract alleviates testis and sperm damages induced by taxane-based chemotherapeutics. *Biomed Pharmacother* 2017; **95**:1284–1294.

Shan J, Cheng W, Zhai DX, Zhang DY, Yao RP, Bai LL, Cai ZL, Liu YH, Yu CQ. Meta-analysis of Chinese traditional medicine Bushen Huoxue prescription for endometriosis treatment. *Evid Based Complement Altern Med* 2017; **2017**:5416423.

Shanmugam MK, Kannaiyan R, Sethi G. Targeting cell signaling and apoptotic pathways by dietary agents: role in the prevention and treatment of cancer. *Nutr Cancer* 2011; **63**:161–173.

Shi M, Shi YL, Li XM, Yang R, Cai CY, Li QS, Ma SC, Ye JH, Lu JL, Liang YR, Zheng XQ. Food-grade encapsulation systems for (–)-epigallocatechin gallate. *Molecules* 2018; **23**:445.

Singh AK, Chakravarty B, Chaudhury K. Nanoparticle-assisted combinatorial therapy for effective treatment of endometriosis. *J Biomed Nanotechnol* 2015; **11**:789–804.

Soleimani V, Delghandi PS, Moallem SA, Karimi G. Safety and toxicity of silymarin, the major constituent of milk thistle extract: an updated review. *Phyther Res* 2019; **33**:1627–1638.

Soliman S, Mohammad MG, El-Keblawy AA, Omar H, Abouleish M, Madkour M, Elnaggar A, Hosni RM. Mechanical and phytochemical protection mechanisms of *Calligonum comosum* in arid deserts. *PLoS One* 2018; **13**:e0192576.

Soo HL, Quah SY, Sulaiman I, Sagineedu SR, Lim JCW, Stanslas J. Advances and challenges in developing andrographolide and its analogues as cancer therapeutic agents. *Drug Discov Today* 2019; **24**:1890–1898.

Su SY, Muo CH, Sung FC, Morisky DE. Reduction of surgery rate in endometriosis patients who take Chinese medicine: a population-based retrospective cohort study. *Complement Ther Med* 2014; **22**:632–639.

Suou K, Taniguchi F, Tagashira Y, Kiyama T, Terakawa N, Harada T. Apigenin inhibits tumor necrosis factor α -induced cell proliferation and prostaglandin E2 synthesis by inactivating NF κ B in endometriotic stromal cells. *Fertil Steril* 2011; **95**:1518–1521.

Swarnakar S, Paul S. Curcumin arrests endometriosis by downregulation of matrix metalloproteinase-9 activity. *Indian J Biochem Biophys* 2009; **46**:59–65.

Szymanowski K. Apoptosis pattern in human endometrium in women with pelvic endometriosis. *Eur J Obstet Gynecol Reprod Biol* 2007; **132**:107–110.

Taguchi A, Koga K, Kawana K, Makabe T, Sue F, Miyashita M, Yoshida M, Urata Y, Izumi G, Tkamura M et al. Resveratrol enhances apoptosis in endometriotic stromal cells. *Am J Reprod Immunol* 2016; **75**:486–492.

Taguchi A, Wada-Hiraike O, Kawana K, Koga K, Yamashita A, Shirane A, Urata Y, Kozuma S, Osuga Y, Fujii T. Resveratrol suppresses inflammatory responses in endometrial stromal cells derived from endometriosis: a possible role of the sirtuin 1 pathway. *J Obstet Gynaecol Res* 2014; **40**:770–778.

Tahmasebi F, Movahedin M, Mazaheri Z. Antioxidant effects of *Calligonum* extract on ovarian tissue of PCO model: an experimental study. *Int J Reprod Biomed (Yazd, Iran)* 2018; **16**:641–648.

Takai E, Taniguchi F, Nakamura K, Uegaki T, Iwabe T, Harada T. Parthenolide reduces cell proliferation and prostaglandin E₂ in human endometriotic stromal cells and inhibits development of endometriosis in the murine model. *Fertil Steril* 2013; **100**:1170–1178.

Takehara M, Ueda M, Yamashita Y, Terai Y, Hung YC, Ueki M. Vascular endothelial growth factor a and C gene expression in endometriosis. *Hum Pathol* 2004; **35**:1369–1375.

Takeuchi A, Koga K, Tokita Y, Matsumoto T, Satake E, Taguchi A, Makabe T, Miyashita M, Takamura M, Harada M et al. The effects of tokishakuyakusan, a traditional Japanese medicine (kampo), ferulic acid and paeoniflorin, on human endometriotic stromal cells and peritoneal macrophages. *J Reprod Immunol* 2020; **139**:103104.

Tan WSD, Liao W, Zhou S, Wong WSF. Is there a future for andrographolide to be an anti-inflammatory drug? Deciphering its major mechanisms of action. *Biochem Pharmacol* 2017; **139**:71–81.

Tang Q, Shang F, Wang X, Yang Y, Chen G, Chen Y, Zhang J, Xu X. Combination use of ferulic acid, ligustrazine and tetrahydropalmatine inhibits the growth of ectopic endometrial tissue: a multi-target therapy for endometriosis rats. *J Ethnopharmacol* 2014; **151**:1218–1225.

Taylor RN, Yu J, Torres PB, Schickendantz AC, Park JK, Mueller MD, Sidell N. Mechanistic and therapeutic implications of angiogenesis in endometriosis. *Reprod Sci* 2009; **16**:140–146.

Teng L, Zu Q, Li G et al. Herbal medicines: challenges in the modern world. Part 3. China and Japan. *Expert Rev Clin Pharmacol* 2016; **9**:1225–1233.

Thomson M, Ali M. Garlic [*Allium sativum*]: a review of its potential use as an anti-cancer agent. *Curr Cancer Drug Targets* 2003; **3**:67–81.

Tomassetti C, Meuleman C, Pexsters A, Mihalyi A, Kyama C, Simsa P, D'Hooghe TM. Endometriosis, recurrent miscarriage and implantation failure: is there an immunological link? *Reprod Biomed Online* 2006; **13**:58–64.

Tsai PJ, Lin YH, Chen JL, Yang SH, Chen YC, Chen HY. Identifying Chinese herbal medicine network for endometriosis: implications from a population-based database in Taiwan. *Evid Based Complement Altern Med* 2017; **2017**:1–11.

Ulbricht C, Brigham A, Burke D, Costa D, Giese N, Iovin R, Grimes Serrano JM, Tanguay-Colucci S, Weissner W, Windsor R. An evidence-based systematic review of acai (*Euterpe oleracea*) by the

natural standard research collaboration. *J Diet Suppl* 2012;9:128–147.

Vassiliadis S, Relakis K, Papageorgiou A, Athanassakis I. Endometriosis and infertility: a multi-cytokine imbalance versus ovulation, fertilization and early embryo development. *Clin Dev Immunol* 2005;12:125–129.

Vinayak M, Maurya AK. Quercetin loaded nanoparticles in targeting cancer: recent development. *Anticancer Agents Med Chem* 2019;19:1560–1576.

Walker OLS, Holloway AC, Raha S. The role of the endocannabinoid system in female reproductive tissues. *J Ovarian Res* 2019;12:3.

Wang CC, Xu H, Man GCW, Zhang T, Chu KO, Chu CY, Cheng JTY, Li G, He YX, Qin L et al. Prodrug of green tea epigallocatechin-3-gallate (Pro-EGCG) as a potent anti-angiogenesis agent for endometriosis in mice. *Angiogenesis* 2013;16:59–69.

Wang D, Liu Y, Han J, Zai D, Ji M, Cheng W, Xu L, Yang L, He M, Ni J et al. Puerarin suppresses invasion and vascularization of endometriosis tissue stimulated by 17 β -estradiol. *PLoS One* 2011;6:e25011.

Wang, R. Chinese University of Hong Kong. Green Tea for Endometriosis. Bethesda, MD, USA: U.S. National Library of Medicine, 2020a. <https://clinicaltrials.gov/ct2/show/NCT02832271> (21 August 2020, date last accessed).

Wang, Z. China Academy of Chinese Medical Sciences. Xuefu-Zhuyu Capsule for the Treatment of "Qizhi Xueyu Zheng" (Qi Stagnation and Blood Stasis Syndrome). Bethesda, MD, USA: U.S. National Library of Medicine, 2020b. <https://clinicaltrials.gov/ct2/show/NCT04218487> (21 August 2020, date last accessed).

Wen XL. Xiao Liu Fang for treatment of 42 cases of uterine fibroids. *Shaanxi J Tradit Chin Med* 2007;28:298–299.

Wieser F, Cohen M, Gaeddert A, Yu J, Burks-Wicks C, Berga S, Taylor R. Evolution of medical treatment for endometriosis: back to the roots? *Hum Reprod Update* 2007;13:487–499.

Wu X, Zhang H, Salmani JMM, Fu R, Chen B. Advances of wogonin, an extract from *Scutellaria baicalensis*, for the treatment of multiple tumors. *Onco Targets Ther* 2016;9:2935–2943.

Xu H, Becker CM, Lui WT, Chu CY, Davis TN, Kung AL, Birsner AE, D'Amato RJ, Wai Man GC, Wang CC. Green tea epigallocatechin-3-gallate inhibits angiogenesis and suppresses vascular endothelial growth factor C/vascular endothelial growth factor receptor 2 expression and signaling in experimental endometriosis *in vivo*. *Fertil Steril* 2011;96:1021–1028.

Xu H, Lui WT, Chu CY, Ng PS, Wang CC, Rogers MS. Anti-angiogenic effects of green tea catechin on an experimental endometriosis mouse model. *Hum Reprod* 2009;24:608–618.

Yavuz S, Aydin N, Celik O, Yilmaz E, Ozerol E, Tanbek K. Resveratrol successfully treats experimental endometriosis through modulation of oxidative stress and lipid peroxidation. *J Cancer Res Ther* 2014;10:324–329.

Yesil-Celiktas O, Sevimli C, Bedir E, Vardar-Sukan F. Inhibitory effects of rosemary extracts, carnosic acid and rosmarinic acid on the growth of various human cancer cell lines. *Plant Foods Hum Nutr* 2010;65:158–163.

Yoshino T, Katayama K, Horiba Y et al. The difference between the two representative kampo formulas for treating dysmenorrhea: an observational study. *Evid Based Complement Altern Med* 2016;2016:3159617.

Yu J, Zhao L, Zhang D, Zhai D, Shen W, Bai L, Liu Y, Cai Z, Li J, Yu C. The effects and possible mechanisms of puerarin to treat endometriosis model rats. *Evid Based Complement Altern Med* 2015;2015:269138.

Yu SS, Zhao J, Zheng WP, Zhao Y. Neuroprotective effect of 4-hydroxybenzyl alcohol against transient focal cerebral ischemia via anti-apoptosis in rats. *Brain Res* 2010;1308:167–175.

Yun-Fei Q, Shu-Juan L, Bin I. Estrogen-independent inhibition of curcumin on formation of endometriotic foci in rats with experimental endometriosis. *Med J West China* 2012;24.

Zhang B, Zhou WJ, Gu CJ, Wu K, Yang HL, Mei J, Yu JJ, Hou XF, Sun JS, Xu FY et al. The ginsenoside PPD exerts anti-endometriosis effects by suppressing estrogen receptor-mediated inhibition of endometrial stromal cell autophagy and NK cell cytotoxicity article. *Cell Death Dis* 2018;9:74.

Zhang L. Pharmacokinetics and drug delivery systems for puerarin, a bioactive flavone from traditional Chinese medicine. *Drug Deliv* 2019;26:860–869.

Zhang L, Zhu L, Wang Y, Jiang Z, Chai X, Zhu Y, Gao X, Qi A. Characterization and quantification of major constituents of Xue Fu Zhu Yu by UPLC-DAD-MS/MS. *J Pharm Biomed Anal* 2012;62:203–209.

Zhang Y, Cao H, Hu YY, Wang H, Zhang CJ. Inhibitory effect of curcumin on angiogenesis in ectopic endometrium of rats with experimental endometriosis. *Int J Mol Med* 2011;27:87–94.

Zhang Y, Cao H, Yu Z, Peng HY, Zhang CJ. Curcumin inhibits endometriosis endometrial cells by reducing estradiol production. *Int J Reprod Biomed* 2013a;11:415–422.

Zhang Z, Cheng X, Gui T, Tao J, Huang M, Zhu L, Luo M, Cao P, Wan G. Wenshen Xiaozheng Tang induces apoptosis and inhibits migration of ectopic endometriotic stromal cells. *J Ethnopharmacol* 2016;194:386–394.

Zhang Z, Hu C, Tang W, Gui T, Qian R, Xing Y, Cao P, Wan G. Therapeutic potential of Wenshen Xiaozheng Tang, a traditional Chinese medicine prescription, for treating endometriosis. *Reprod Sci* 2013b;20:1215–1223.

Zhao J, Liu L, Zhang Y, Wan Y, Hong Z. the herbal mixture Xiao-Chai-Hu Tang (XCHT) induces apoptosis of human hepatocellular carcinoma HuH7 cells *in vitro* and *in vivo*. *Afr J Tradit Complement Altern Med* 2017;14:231–241.

Zhao RH, Hao ZP, Zhang Y, Lian FM, Sun WW, Liu Y, Wang R, Long L, Cheng L, Ding YF et al. Controlling the recurrence of pelvic endometriosis after a conservative operation: comparison between Chinese herbal medicine and western medicine. *Chin J Integr Med* 2013;19:820–825.

Zheng Y, Liu X, Guo SW. Therapeutic potential of andrographolide for treating endometriosis. *Hum Reprod* 2012;27:1300–1313.

Zhong YC, Zhou XF, Hou CM, Li WP. Effect of Danefukang on symptoms and biomarkers in women with endometriosis. *Taiwan J Obstet Gynecol* 2019;58:218–222.

Zhou H, Zhang Q, Qi C. Impact of a Chinese medicinal formula, Xiao Liu Fang, on the "3A" ability of endometrial stromal cells in patients with endometriosis. *Mol Ther Oncolytics* 2019;12:1–8.

Zhou J, Ding ZM, Hardiman PJ. Understanding the role of Gui-Zhi-Fu-Ling-capsules (Chinese medicine) for treatment of endometriosis in the rat model: using NMR based metabolomics. *Evid Based Complement Altern Med* 2018;2018:9864963.

Zhou X, Seto SW, Chang D, Kiat H, Razmovski-Naumovski V, Chan K, Bensoussan A. Synergistic effects of Chinese herbal medicine: a comprehensive review of methodology and current research. *Front Pharmacol* 2016;7:201.

Zhu G, Jiang C, Yan X, Zhao S, Xu D, Cao Y. Shaofu Zhuyu decoction regresses endometriotic lesions in a rat model. *Evid Based Complement Altern Med* 2018;2018:3927096.

Zondervan KT, Becker CM, Koga K, Missmer SA, Taylor RN, Viganò P. Endometriosis. *Nat Rev Dis Prim* 2018;4:9.

Zou J, Guan Z, Zhang WY, Xiao W, Li YL. Beneficial effects of the Chinese herbal medicine Sanjie Zhentong capsule on experimental endometriosis in rats. *Chin J Nat Med* 2013;11:666–672.